TECHNICAL DATA SHEET

AlphaLISA® Research Reagents

AlphaLISA Human FCGR1/CD64 Binding Kit

Product number: AL3081C/F

Lot number: 2988554

Manufacturing date: March 2, 2022

Research Use Only. Not for use in diagnostic procedures.

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Product Information

Application: This kit is designed for the detection of binding between FCGR1 (CD64) and human IgG Fc fragment using a homogeneous AlphaLISA assay (no wash steps). This assay can facilitate the design and development of antibody therapetics by using competitive binding.

Sensitivity: IC_{50} 0.59 µg/mL (average, with IgG whole molecule)

Signal to background ratio: 157 (average)

Kit contents: The kit contains 4 components: Human IgG Fc fragment conjugated Acceptor beads, Streptavidin-coated Donor beads, Biotinylated human FCGR1, and AlphaLISA HiBlock buffer.

Storage: Store kit in the dark at 4 ˚C.

Stability: This kit is stable for at least 6 months from the manufacturing date when stored in its original packaging and the recommended storage conditions. After reconstitution, store unused protein in -20 °C for 3 months. Avoid multiple freeze/thaw cycles.

Quality Control

Lot to lot consistency is confirmed in an AlphaLISA assay. Maximum and minimum signals were measured on the EnVision Multilabel Plate Reader with Alpha option using the protocol described in this technical data sheet. We certify that these results meet our quality release criteria. Maximum and minimum counts may vary between bead lots and the instrument used**.**

IC_{50}:   0.64 µg/mL
S/B   244

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Analyte of Interest

The Fc-Gamma Receptors (FCGRs) are members of immunoglobulin superfamily and play a critical role in the function of therapeutic antibodies. FCGRs are divided into three classes Fc-Gamma Receptor 1 (CD64), FCGR1; Fc-Gamma Receptor 2 (CD32), FCGR2 and Fc-Gamma receptor 3 (CD16), FCGR3.

FCGR1 is a high affinity Fc-gamma receptor, also known as FCGR1A and CD64. FCGR1 is a transmembrane protein with three extracellular Ig-like domains, and it delivers an activating signal via the associated Fc R gamma accessory chain. FCGR1 permits the triggering of effector responses at low IgG concentrations typical of early immune responses.

Description of the AlphaLISA Assay

The AlphaLISA detection of FCGR1 and IgG Fc fragment binding uses IgG Fc AlphaLISA® acceptor beads to capture the human FCGR1 and Streptavidin-coated donor beads to capture the biotinylated human FCGR1. Donor beads and acceptor beads come into proximity through IgG Fc fragment binding to FCGR1. Excitation of the Donor beads provokes the release of singlet oxygen that triggers a cascade of energy transfer reactions in the Acceptor beads, resulting in a sharp peak of light emission at 615 nm (Figure 1).
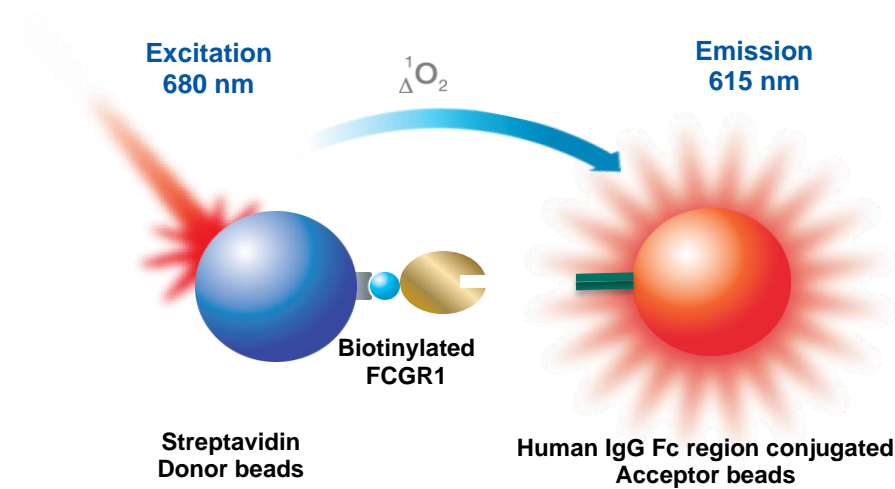

Excitation 680 nm

$^1_\Delta O_2$

Emission 615 nm

Biotinylated FCGR1

Streptavidin Donor beads

Human IgG Fc region conjugated Acceptor beads

Figure 1. AlphaLISA assay principle.

Precautions

- The AlphaScreen® Donor beads are light-sensitive. All the other assay reagents can be used under normal light conditions. All Alpha assays using the Donor beads should be performed under subdued laboratory lighting (< 100 lux). Green filters (LEE 090 filters (preferred) or Roscolux filters #389 from Rosco) can be applied to light fixtures.

Kit Content: Reagents and Materials

Kit components	AL3081C (500 assay points)**	AL3081F (5000 assay points)**

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AlphaLISA Human IgG Fc fragment Acceptor beads stored in PBS, 0.05% Kathon, pH 7.2	80 µL @ 5 mg/mL (1 brown tube, <u>white</u> cap)	800 µL @ 5 mg/mL (1 brown tube, <u>white</u> cap)
Streptavidin (SA)-coated Donor beads stored in 25 mM HEPES, 100 mM NaCl, 0.05% Kathon, pH 7.4	80 µL @ 5 mg/mL (1 brown tube, <u>black</u> cap)	800 µL @ 5 mg/mL (1 brown tube, <u>black</u> cap)
Human CD64 / FCGR1A Protein (His Tag), Biotinylated (lyophilized solid***)	1 µg (1 tube, <u>clear</u> cap)	10 x 1 µg (10 tubes, <u>clear</u> caps)
AlphaLISA HiBlock Buffer (10X)*	10 mL, 1 large bottle	100 mL, 1 large bottle

* Extra HiBlock buffer can be ordered separately (cat # AL004 C: 10 mL, cat # AL004F: 100 mL).

** The number of assay points is based on an assay volume of 40 µL in 96-well assay plates using the kit components at the recommended concentrations.

*** After reconstitution, aliquot and store unused protein at -20 °C for 3 months. Avoid multiple freeze/thaw cycles.

Additional Reagents and Materials

The following items are recommended for the assays:

Item	Supplier	Catalog number
½ AlphaPlate-96, white	PerkinElmer	6005560 (50/box) 6005569 (200/box)
TopSeal™-A Plus Adhesive Sealing Film	PerkinElmer	6050185
EnSpire® or EnVision® Multilabel Alpha Reader	PerkinElmer	Please consult our website

The following reagents might be required for particular applications:

Item	Supplier	Catalog number
IgG1 Human Plasma	Athens Research Technology	16-16-090707-1
IgG2 Human Plasma	Athens Research Technology	16-16-090707-2
IgG3 Human Plasma	Athens Research Technology	16-16-090707-3
IgG4 Human Plasma	Athens Research Technology	16-16-090707-4
ChromPure Human IgG F(ab')$_2$ Fragment	JacksonImmunoResearch	009-000-006
ChromPure Human IgG Fc Fragment	JacksonImmunoResearch	009-000-008

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ChromPure Human IgG,whole molecule	JacksonImmunoResearch	009-000-003
Anti-human CD64 antibody	Bio-Rad Laboratories	MCA756GT

Recommendations

- The volume indicated on each tube is guaranteed for single pipetting. Multiple pipetting of the reagents may reduce the theoretical amount left in the tube. To minimize loss when pipetting beads, it is preferable not to prewet the tip.

- Centrifuge quickly all tubes before use to improve recovery of content (2 000 ×g, 10-15 sec). Resuspend all reagents by gentle mixing before use.

- Use Milli-Q® grade H_2O to dilute 10X HiBlock Buffer 1.

- When reagents are added in the microplate, make sure the liquids are at the bottom of the well by tapping or swirling the plate gently on a smooth surface.

- Small volumes may be prone to evaporation. It is recommended to cover microplates with TopSeal™-A Plus Adhesive Sealing Film to reduce evaporation during incubation with the Alpha beads. Microplates can be read with the TopSeal-A Film.

- The AlphaLISA signal is detected with an EnVision Multilabel Reader equipped with the ALPHA option using the AlphaScreen standard settings (e.g. Total Measurement Time: 550 ms, Laser 680 nm Excitation Time: 180 ms, Mirror: D640as (barcode 444), Emission Filter: M570w (barcode 224), Center Wavelength 570 nm, Bandwidth 100 nm, Transmittance 75%).

- AlphaLISA signal will vary with temperature and incubation time. For consistent results, identical incubation time and temperature should be used for each plate.

Competition Assay Protocol

- Assay specificity can be demonstrated by competing the binding of human FCGR1 with all subclasses of human IgG or comparing the binding with human IgG fragments.

The competition assay described below is an example for determining IC$_{50}$ of human IgG subclass competitive binding to human FCGR1 in 40 µL final assay volume (96 wells, duplicate determinations) by AlphaLISA technology. This protocol can test 4 antibodies in one 96 wells. If a different amount of samples are tested, the volumes of all reagents have to be adjusted accordingly. The protocol is provided for information only. As needed, the number of replicates or the range of concentrations covered can be modified.

1. Preparation of 1X HiBlock Buffer (for 10 mL)
    Add 1 mL of 10X HiBlock Buffer and 9 mL of MilliQ water.

2. Preparation of serial dilution of human IgG subclasses

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Prepare serial dilutions of IgG in 1X HiBlock buffer as follows. Change tips between each dilution:

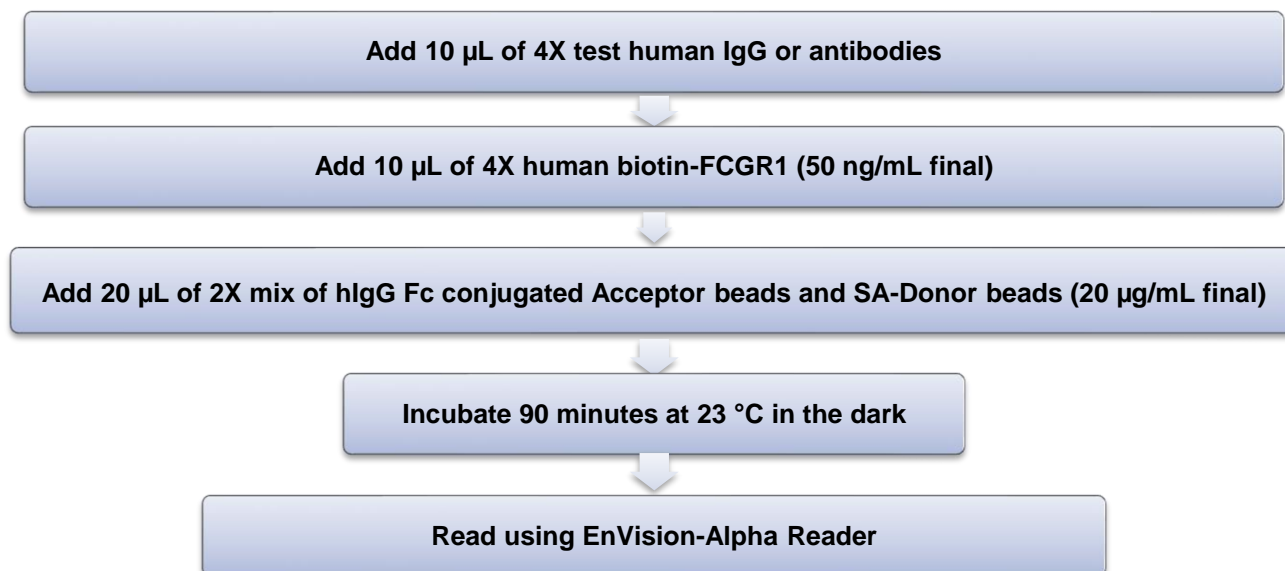| Tube	Volume of IgG	Volume of 1X buffer	[IgG] (g/mL) (4X)	[IgG] (g/mL) (1X)
A	1.2 mg/mL stock	0	1.20E-03	3.00E-04
B	30 µL of tube A	60 µL	4.00E-04	1.00E-04
C	30 µL of tube B	70 µL	1.20E-04	3.00E-05
D	30 µL of tube C	60 µL	4.00E-05	1.00E-05
E	30 µL of tube D	70 µL	1.20E-05	3.00E-06
F	30 µL of tube E	60 µL	4.00E-06	1.00E-06
G	30 µL of tube F	70 µL	1.20E-06	3.00E-07
H	30 µL of tube G	60 µL	4.00E-07	1.00E-07
I	30 µL of tube H	70 µL	1.20E-07	3.00E-08
J	30 µL of tube I	60 µL	4.00E-08	1.00E-08
K	30 µL of tube J	70 µL	1.20E-08	3.00E-09
L		60 µL	0	0

3. Preparation of 4X human FCGR1 (200 ng/mL)
   a. Spin the vial containing 1 µg lyophilized protein briefly in microfuge and reconstitute it with 100 µL sterile distilled water to make 10 µg/mL stock concentration of human FCGR1.

   b. Add the 20 µL of 10 µg/mL human FCGR1 into a new tube containing 980 µL 1X HiBlock Buffer to make 200 nM FCGR1. After reconstitution, aliquot and store unused protein at -20 °C for 3 months. Avoid multiple freeze/thaw cycles.

   c. Prepare just before use.

4. Preparation of 2X mix of human IgG Fc Conjugated Acceptor Beads (40 µg/mL) and Streptavidin (SA) Donor Beads (40 µg/mL).
   a. Add 16 µL of 5 mg/mL human IgG Fc conjugated Acceptor beads and 16 µL of 5 mg/mL SA-Donor beads into 1968 µL 1X HiBlock buffer.

   b. Keep the beads under subdued laboratory lighting and prepare just before use.

5. In a ½ AreaPlate (96 wells):

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Add 10 µL of 4X test human IgG or antibodies

Add 10 µL of 4X human biotin-FCGR1 (50 ng/mL final)

Add 20 µL of 2X mix of hIgG Fc conjugated Acceptor beads and SA-Donor beads (20 µg/mL final)

Incubate 90 minutes at 23 °C in the dark
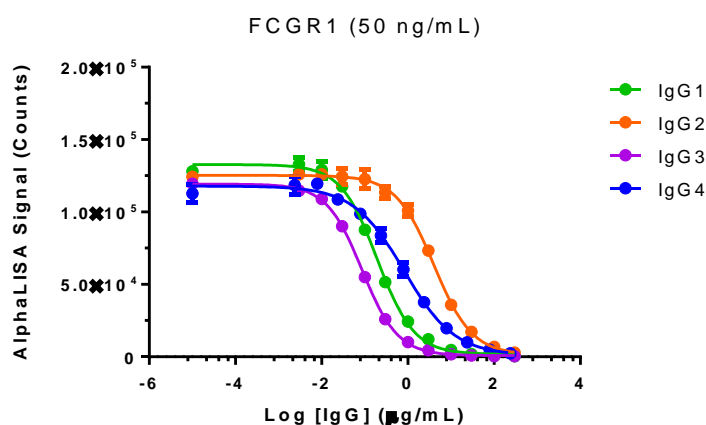
Read using EnVision-Alpha Reader

Typical competitive binding Data:

Figure 2. Human IgG subclasses competitive bind to FCGR1. The IC$_{50}$ values were 0.196, 4.036, 0.087 and 0.815 µg/mL for IgG1, IgG2, IgG3 and IgG4 respectively and were calculated by using nonlinear regression fitting with GraphPad Prism 6. Each IgG subclass has gone through a zeba column (ThermoFisher, Cat. no. 89882) for a buffer exchange with PBS before testing. The concentrations of IgGs were measured with NanoDrop (E 1%=13.6).

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FCGR1 (50 ng/mL)
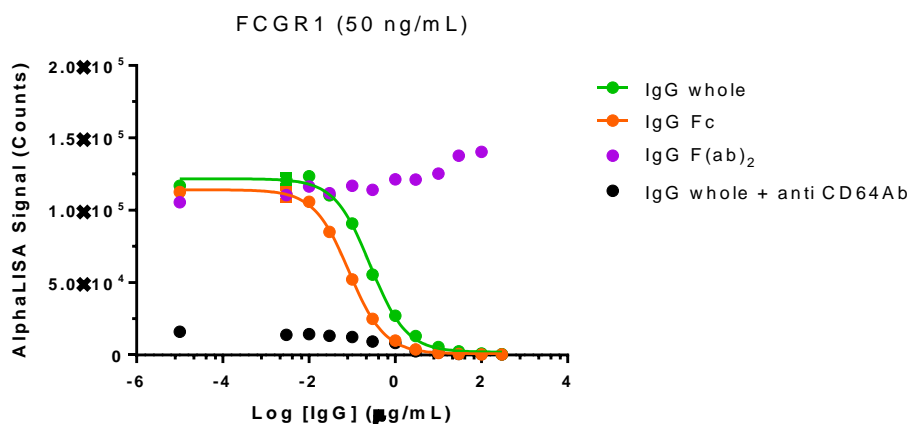
Figure 3. Human IgG fragments competitive bind to FCGR1. Black points showed human IgG whole molecule which was pre-incubated with anti-human CD64 antibody for 10 minutes at room temperature as a negative control. The IC_{50} values were 0.266 and 0.086 μg/mL for IgG whole molecule and IgG Fc fragment respectively and were calculated by using nonlinear regression fitting with GraphPad Prism 6. The IC_{50} was not measurable for IgG F(ab)$_2$.

Troubleshooting Guide

You will find below recommendations for common situations that you might encounter with your AlphaLISA detection assay. If further assistance is needed, do not hesitate to contact our technical support team for assistance.

Issue	Recommendations and Comments
High background signal	• Buffer is not freshly made. Make new. • Incubation time is longer than recommended range.
Low AlphaLISA signal	• Optimize EnVision with Plate format.
High variation between replicates or low Z' values	• Make sure that reagents are at the bottom of the well by tapping or swirling the plate gently on a smooth surface after each addition.

You will find detailed recommendations for common situations you might encounter with your AlphaLISA Assay kit at:

http://www.perkinelmer.com/in/resources/technicalresources/applicationsupportknowledgebase/alphalisa-alphascreen-no-washassays/alpha_troubleshoot.xhtml

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