DNA 12K Assay User Guide

For LabChip GX Touch/GXII Touch

CE

P/N CLS140167 Rev. D

Publication Date: August 24, 2018

Contents

Specifications

Assay Specifications

Table 1. Assay Specifications

Sizing Range	100-12000 bp
Sizing Resolution[a]	10% from 150 - 1000 bp 15% from 1000 - 2000 bp 20% from 2000 - 8000 bp 25% from 100 - 150 bp, 8000 - 12000 bp
Sizing Accuracy	± 10%
Sizing Precision	5% CV
Linear Concentration Range	0.25 - 50 ng/µL per fragment
Sensitivity	0.25 ng/µL
Maximum Total DNA Concentration	60 ng/µL total, 50 ng/µL per fragment
Quantitation Accuracy	± 40% or ± 1 ng/µL, whichever is greater
Quantitation Precision	20% CV from 100 - 5000 bp, 25% CV from 5000 - 12000 bp
Number of Samples per Chip Prep	400 samples (four 96-well plates or one 384-well plate)

a. Resolution is defined as half height or better separation of two peaks. Actual separation performance can depend on the sample and application. Peaks that are resolved less than half height can still be accurately identified by the system software.

Sample Conditions

Table 2. Sample Conditions

Additives	PerkinElmer recommends that BSA and detergents exceeding 0.05 mg/mL and 0.01% (v/v) respectively in concentration not be used. Higher concentrations can result in chip failure. In addition, non-aqueous solvents are not compatible with DNA LabChip protocols.
Particulates	All sample plates should be spun down prior to analysis. All buffers should be filtered with a 0.22 µm cellulose acetate filter.
Salt Concentration	Total salt concentration must not exceed 125 mM.
Plasmids	Plasmid concentration in samples must be below 20 ng/µL. Please note that although the DNA Assays cannot analyze plasmids, the presence of plasmids above 20 ng/µL can interfere with assay results

Kit Contents

Storage: Store chips and reagents refrigerated at 2-8°C until next use. If using the chip again within 24 hours it may be left at room temperature. Allowing the chip wells to dry may lead to changes in chip performance.

Table 3. Reagent Kit Contents, PN 760569

Reagent	Vial	Quantity
DNA Dye Concentrate	Blue	1 vial, 0.08 mL
DNA Chip Storage Buffer	White	5 vials, 1.8 mL each
DNA 12K Gel Matrix	Red	3 vials, 1.6 mL each
DNA 12K Ladder, 10K	Yellow	1 vial, 0.15 mL
DNA 12K Marker	Green	1 vial, 1.5 mL

Table 4. Consumable Items

Item	Supplier and Catalog Number	Quantity
Spin Filters	Costar®, Cat. # 8160	8
Detection Window Cleaning Cloth	VWR®, Cat. # 21912-046	1
Swab	ITW Texwipe®, Cat. # TX758B	3

Table 5. DNA LabChips

Item	Catalog Number
DNA Extended Range Chip (12K) for use with GX Touch/GXII Touch HT	Cat. # 760517
DNA Extended Range Chip (12K) for use with GX Touch/GXII Touch 24	Cat. # CLS138948

Safety and Usage

Safety Warnings and Precautions

WARNING!

For Research Use Only. Not recommended or intended for diagnosis of disease in humans or animals. Do not use internally or externally in humans or animals.

CAUTION

We recommend that this product and components be handled only by those who have been trained in laboratory techniques and that it is used in accordance with the principles of good laboratory practice. As all chemicals should be considered as potentially hazardous, it is advisable when handling chemical reagents to wear suitable protective clothing, such as laboratory overalls, safety glasses, and gloves. Care should be taken to avoid contact with skin or eyes. In case of contact with skin or eyes, wash immediately with water

WARNING!

Dye Concentrate contains DMSO. S24/25: Avoid contact with skin and eyes.

Usage

The DNA 12K assay is for use with LabChip GX Touch/GXII Touch instruments. The LabChip GX Touch/GXII Touch instruments are for research use only and not for use in diagnostic procedures.

Preparation Procedures

Additional Items Required

- 18 megohm, 0.22-µm filtered water (Milli-Q® or equivalent).
- 70% isopropanol solution in DI water.
- Bio-Rad Hard-Shell® 384-well Skirted PCR Plates, Cat. # HSP-38XX (recommended)
- PerkinElmer Hard-Shell thin-wall 96-well skirted PCR plate (blue), Cat. # 6008870 (recommended)

Note: *Allow the chip and reagents to equilibrate to room temperature for at least 30 minutes before use.*

Preparing the Gel-Dye Solution

Notes: *The Dye Concentrate contains DMSO and must be thawed completely before use.*

The dye is light sensitive. Do not expose Dye or Gel-Dye to light for any length of time. Keep prepared Gel-Dye in the dark.

1 Vortex the thawed DNA Dye Concentrate (blue cap 🔵) for 10 -15 seconds before use.

2 Transfer 1.0 mL of DNA Gel Matrix (red cap 🔴) using a reverse pipetting technique to the centrifuge tube provided with the reagent kit. Add 12.5 µL of DNA Dye Concentrate (blue cap 🔵) to the centrifuge tube.

3 Vortex the solution until it is well mixed and spin down for a few seconds.

4 Transfer the mixture into two spin filters (approximately 550 µL each).

5 Centrifuge at 9200 rcf for 7.5 minutes at room temperature.

6 Discard filters, label and date the tubes

7 Store in the dark at 2-8°C. Use within 3 weeks.

Preparing the DNA Samples and DNA Ladder

Notes: *DNA Ladder should be prepared in the same buffer as your DNA samples. A buffer mismatch between sample and ladder may lead to inaccurate quantitation and sizing.*

DNA sample buffer solution is the user's DNA buffer such as PCR buffer, etc.

1. In the provided 0.2 mL Ladder Tube, add 12 µL of DNA Ladder (yellow cap) to 108 µL of 1X DNA sample buffer solution. Mix thoroughly by pipetting the solution up and down a few times. Avoid creating air bubbles. Ensure there are no air bubbles in the Ladder Tube.
2. Insert the Ladder Tube into the ladder slot on the LabChip GX Touch/GXII Touch instrument.
3. Recommended sample volumes are 25 µL for a 384-well plate or 40 µL for a 96-well plate.

Preparing the Buffer Tube

1. Add 750 µL of 1X DNA Sample Buffer solution to the 0.75 mL Buffer Tube provided with the reagent kit. Ensure there are no air bubbles in the Buffer Tube.
2. Insert the Buffer Tube into the buffer slot on the LabChip GX Touch/GXII Touch instrument.

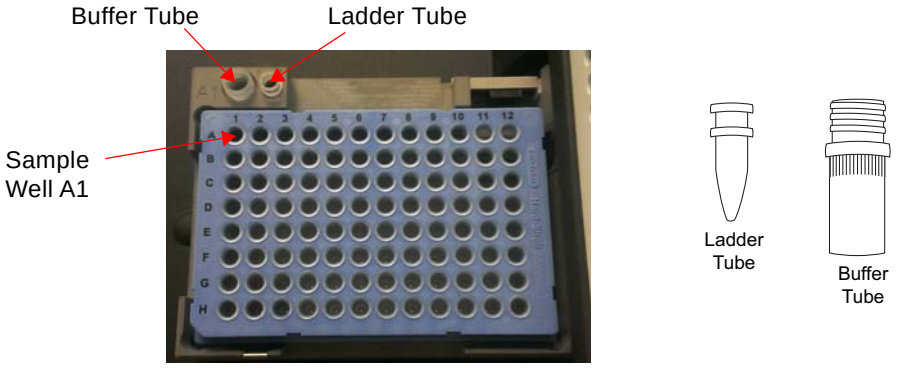

Figure 1. Locations of the Buffer Tube and Ladder Tube in the GXII Touch instrument.

Preparing the Chip

1. Allow the chip to equilibrate to room temperature for at least 30 minutes before use.

2 Use a pipette tip attached to a vacuum line to thoroughly aspirate all fluid from the chip wells (see Figure 2). For more details on how to set up a vacuum line see page 29.

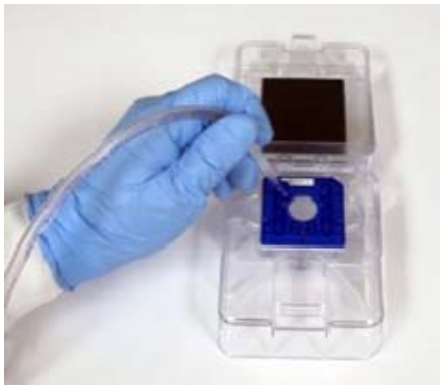

Figure 2. Using a vacuum to aspirate the chip wells is more effective than using a pipette.

3 Rinse and completely aspirate each active chip well (1, 3, 4, 7, 8, and 10) twice with water (Milli-Q® or equivalent). Do not allow active wells to remain dry.

4 If any water spills onto the top and bottom chip surfaces during rinsing, aspirate using the vacuum line. DO NOT run the tip over the central region of the detection window. Use the provided Detection Window Cleaning Cloth dampened in water (Milli-Q® or equivalent) or alcohol to clean chip detection window as needed.

5 Using a reverse pipetting technique, add 75 µL Gel-Dye solution to chip wells 3, 7 and 8, and 120 µL to well 10 as shown in Figure 3.

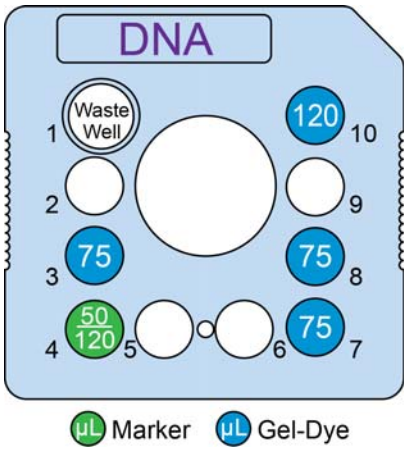

Figure 3. Reagent placement.

6 Add DNA Marker (green cap ●) to chip well 4. Use 50 µL DNA Marker for a 96-well plate and 120 µL DNA Marker for a 384-well plate or multiple 96-well plates as shown in Figure 3.

Note: *The marker well may need to be replenished if the chip is in idle mode on the instrument for an extended period of time.*

7 Make sure the rims of the chip wells are clean and dry.

8 **IMPORTANT:** Ensure chip well 1 (waste well) is empty before placing the chip into the instrument.

Inserting a Chip into the LabChip GX Touch/GXII Touch Instrument

1 Check that the sample plate, Buffer Tube, and Ladder Tube are properly placed on the instrument.

2 Remove the chip from the chip storage container and inspect the chip window. Clean BOTH sides of the chip window with the PerkinElmer-supplied clean-room cloth dampened with a 70% isopropanol solution in DI water.

3 Touch the *Unload Chip* button on the *Home* screen.

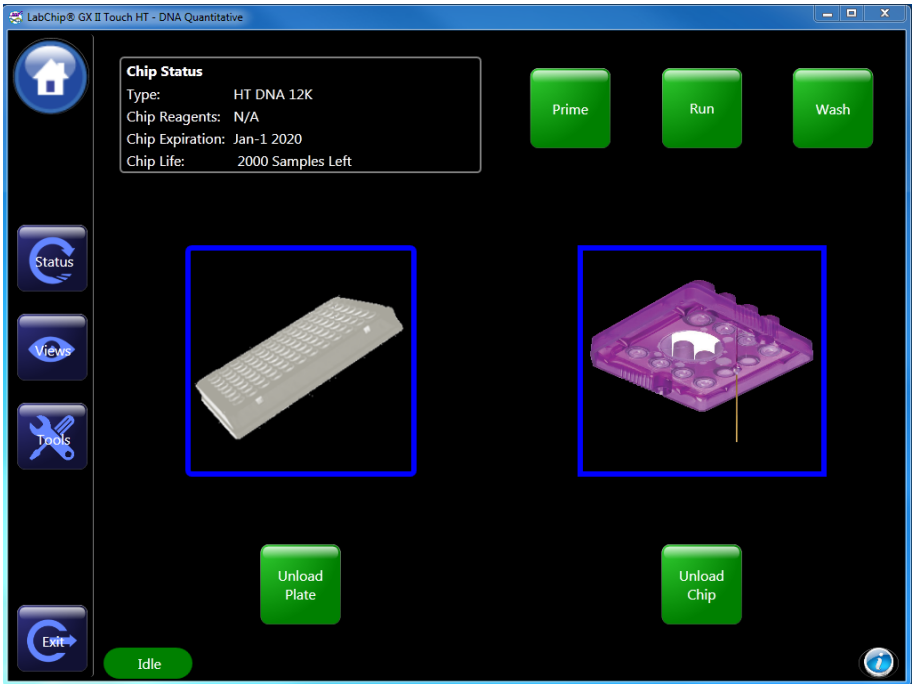

Figure 4. *Home* screen.

4 Insert the chip into the LabChip GX Touch/GXII Touch instrument (Figure 5) and close the chip door securely.

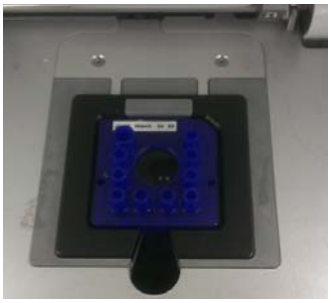

Figure 5. Chip in the LabChip GX Touch/GXII Touch instrument.

5 Touch the *Load Plate* button on the *Home* screen (Figure 4) to retract the sample plate and send the sipper to the Buffer Tube.

Note: *Do not keep the chip door open for any length of time. Dye is sensitive to light and can be photobleached.*

6 The *Assay Choice* window will appear (Figure 6). Touch the desired assay and then touch *OK*.

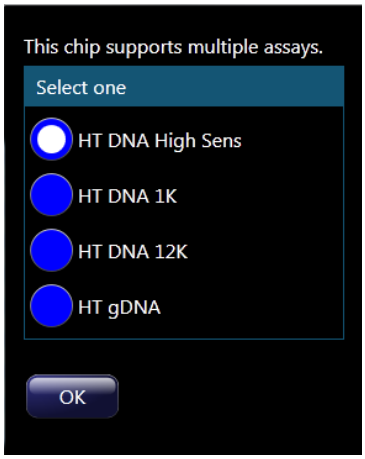

Figure 6. *Assay Choice* menu.

Notes: *The chip may be run with multiple assays, but only one assay type should be run on the chip.*

Be sure to periodically clean the O-rings on the top plate of the chip interface on the LabChip GX Touch/GXII Touch. Use the provided lint-free swab dampened with water to clean the O-rings using a circular motion. Allow the O-rings to dry before inserting a chip.

Running the Assay

Note: *Chips can be primed independently from running assays. Touch the Prime button on the Home screen.* ***Make sure the Buffer Tube is placed on the instrument.***

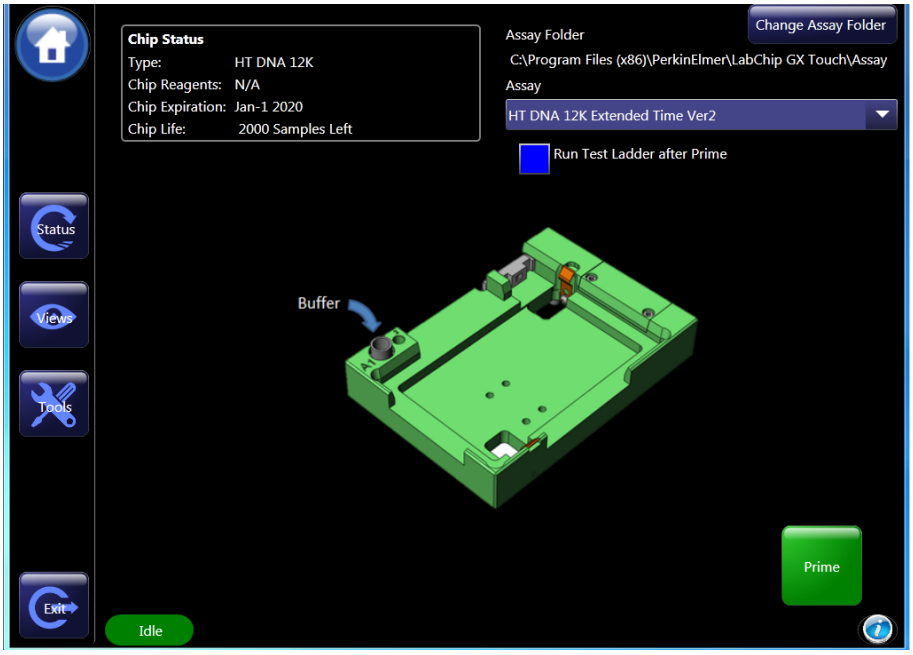

Figure 7. Chip priming screen.

1 Touch the *Run* button (see Figure 7).

2 Select the appropriate assay type (see m). For DNA 12K assays the appropriate assay types are:

- **DNA 12K:** For sizing of DNA fragments in the 100 to 12000 bp range.
- **DNA 12K Extended Time:** To be used only if peaks are cut off using the standard DNA 12K script (occurs in some high salt sample buffers).

Figure 8. *Assay Choice* window.

3 Select the plate name, well pattern, and whether to read wells in columns or rows. Select number of times each well is sampled under *Adv. Settings* (Figure 9). Touch the *green arrow* button.

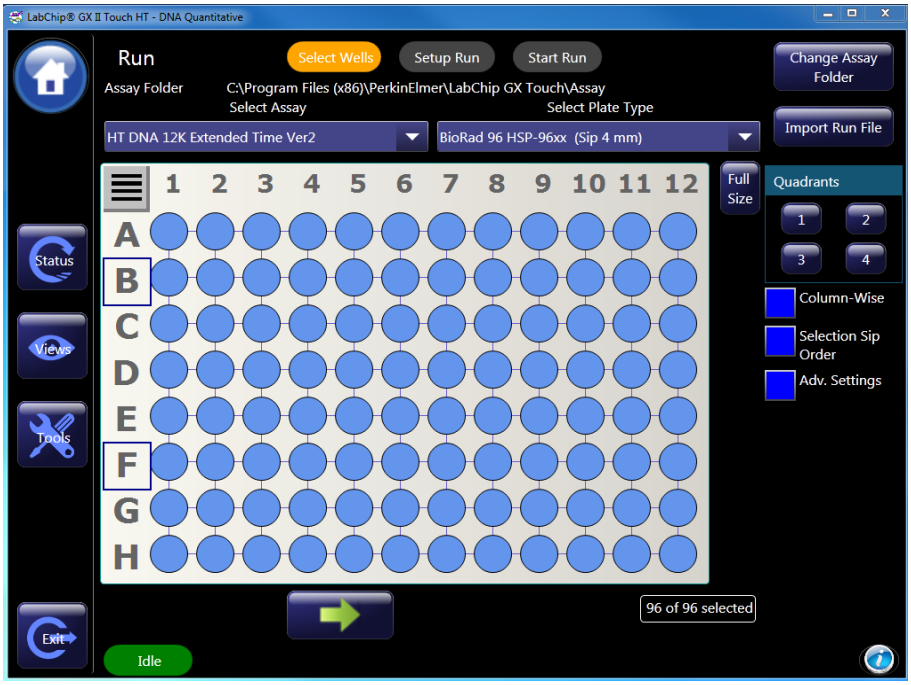

Figure 9. Selecting wells.

4 In the *Setup Run* tab, select the operator name, the option to read barcode, the destination of the file, the inclusion of sample names, expected peaks, and excluded peaks and the filename convention. Select *Auto Export* to export results tables automatically (Figure 10). Touch the *green arrow* button.

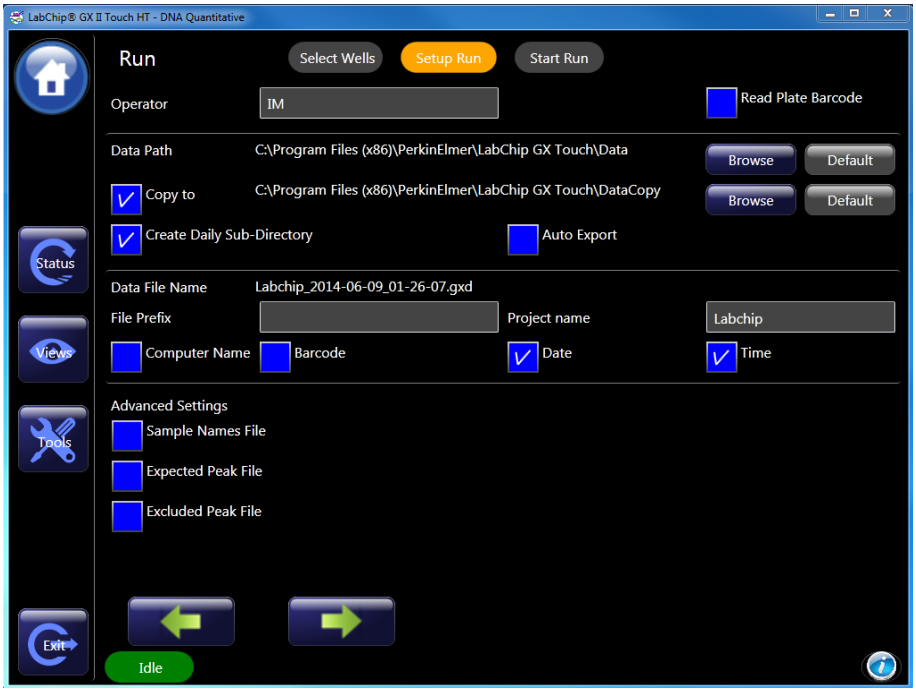

Figure 10. Run setup screen.

5 Touch *Start* to begin the run (Figure 11).

Figure 11. Starting a run.

Cleaning and Storing the Chip

After use, the chip must be cleaned and stored in the chip container.

1. Place the chip into the plastic storage container. The sipper should be submerged in the fluid reservoir.
2. Remove the reagents from each well of the chip using vacuum.
3. Each active well (1, 3, 4, 7, 8, and 10) should be rinsed and aspirated twice with water (Milli-Q® or equivalent).
4. Add 100 µL of Storage Buffer (white cap ◯) to the active wells.
5. Place the chip in the LabChip GX Touch/GXII Touch instrument. Ensure that a Buffer Tube with 750 µL of water (Milli-Q® or equivalent) is in the buffer slot.
6. Touch the *Wash* button in the upper right corner in the *Home* Screen. The *Wash* screen opens (Figure 12).

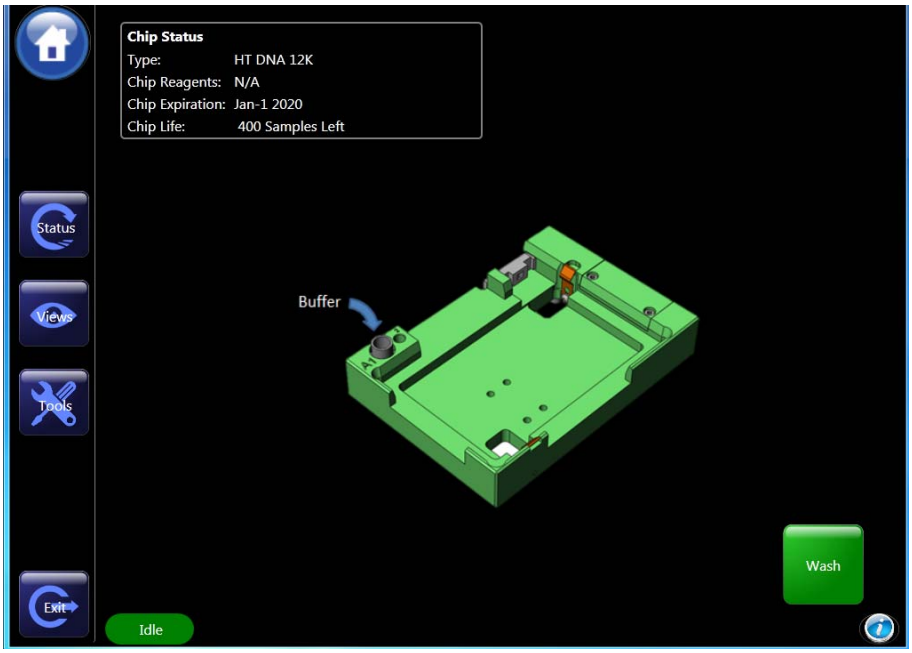

Figure 12. *Wash* screen.

7. Remove the chip from the instrument and place it in the plastic storage container.
8. Add an additional 50 µL of DNA Storage Buffer to well 1.

9 Cover the wells with Parafilm® to prevent evaporation and store at 2-8°C. If using the chip again within 24 hours it may be left at room temperature. Allowing the chip wells to dry may lead to changes in chip performance.

Chip Cartridge Cleaning

1 Daily

a Inspect the inside of the chip cartridge and O-rings for debris.

b Use the provided lint-free swab dampened with water (Milli-Q® or equivalent) to clean the O-rings using a circular motion. If the O-rings stick to the chip or a pressure leak is detected, perform the more extensive monthly cleaning procedure.

2 Monthly

a To reduce pressure leaks at the chip interface, clean the O-rings frequently. Remove the O-rings from the top plate of the chip interface on the LabChip GX Touch/GXII instrument. Soak O-rings in water (Milli-Q® or equivalent) for a few minutes. Clean the O-ring faces by rubbing between two fingers. Wear gloves.

b To reduce the occurrence of current leaks, clean the chip interface frequently. Clean the top plate of the chip interface using the provided lint free swab dampened with water (Milli-Q® or equivalent).

c Allow the O-rings and chip interface to air dry. Reinsert the O-rings into the chip cartridge.

Results

DNA 12K Ladder Result

The electropherogram of a typical DNA 12K ladder is shown in Figure 13. Between the upper and lower markers, peaks in order of increasing migration time correspond to ladder fragments of 100, 300, 500, 700, 1100, 1900, 2900, 4900, 7000 and 10000 bp.

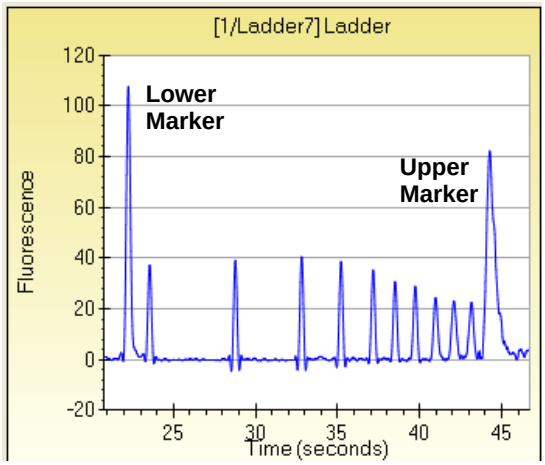

Figure 13. DNA 12K ladder electropherogram.

Troubleshooting

Note: *Some of the data examples shown in this section were generated with assays other than the assay described in this user guide.*

Symptom: No ladder or sample peaks but marker peaks detected.

Note: *The lower marker peak height will most likely be greater than normal height.*

Possible causes:

1 Air bubble in sipper introduced during chip priming.

What to do:

1 Reprime the chip. See "LabChip Kit Essential Practices" on page 23 for instructions on how to reprime the chip.

Symptom: Missing sample, ladder *and* marker peaks.

Possible causes:

1 Clog in sipper or marker channel of chip.

What to do:

1 Reprime the chip. See "LabChip Kit Essential Practices" on page 23 for instructions on how to reprime the chip.

Symptom: Ladder detected but no sample peaks.

Possible causes:

1 The sipper is not reaching the sample due to low sample volume in the well of the plate.

2 If the missing sample peaks occurred only in a few wells of the plate, check those wells for air bubbles.

3 The sipper is not reaching the sample due to an incorrect capillary height setting or incorrect plate definition.

4 If the plate has been uncovered for some time, sample evaporation might have occurred.

5 Debris from the sample or sample prep is clogging the sipper.

What to do:

1 Add more sample to the well.

2 Manually insert a larger volume pipette tip (~100 µL) into the sample well and dislodge the bubble. Rerun these sample wells.

3 Check the plate definitions.

4 Check the sample wells, especially around the edge of the plate where evaporation is fastest, and make a fresh plate if volumes are low.

5 If you suspect there may be debris in your samples, spin the sample plate down in a centrifuge (e.g. 3000 rcf for 5 minutes). Unclog the sipper by repriming the chip. See "LabChip Kit Essential Practices" on page 23 for instructions on how to reprime the chip.

Symptom: No ladder peaks but sample peaks and marker peaks are present.

Possible causes:

1 Low or no ladder volume in the Ladder Tube.

What to do:

1 Add more ladder to the Ladder Tube and restart the run. Recommended standard ladder volume is 120 µL (minimum volume is 100 µL).

Symptom: No marker peaks but sample peaks are present.

Possible causes:

1 No marker added to chip well 4.

2 If there is marker solution in chip well 4, the problem may be due to a marker channel clog.

What to do:

1 This may be due to not filling marker well or chip remaining idle on instrument for extended period of time. Add or replenish the marker solution in the chip using the following procedure:

- Touch the *Unload Chip* button on the Home screen to open the chip door.
- Return the chip to the chip container ensuring the sipper is immersed in fluid.
- Thoroughly aspirate all fluid from chip well 4 using a vacuum line.
- Ensure that chip well 4 is rinsed and completely aspirated twice with water (Milli-Q® or equivalent).
- Add Marker Solution (green cap) to chip well 4.

- Reinsert the chip back into the instrument.
- Restart the run.

2 Perform a marker channel unclogging procedure by repriming the chip. See “LabChip Kit Essential Practices” on page 23 for instructions on how to reprime the chip.

Symptom: Ladder traces show up in the lanes following the ladders (delayed sip).

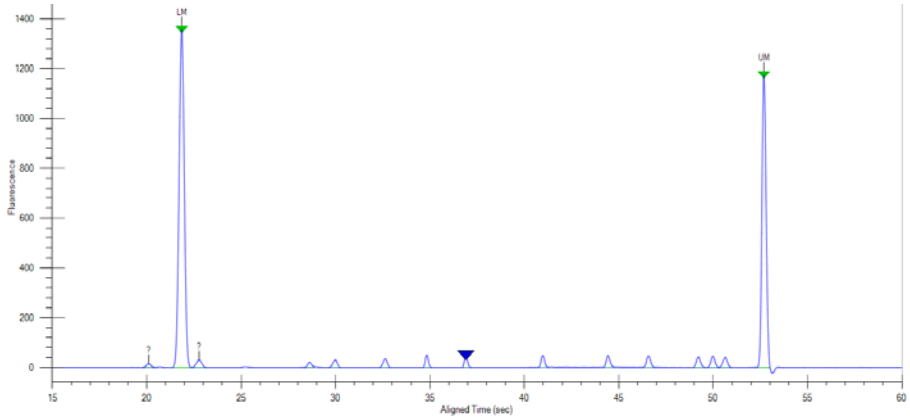

Figure 14. Small ladder peaks in sample well caused by delayed sip.

Possible causes:

1 Separation channel overloaded with sample.

2 Partial clog in the separation channel.

What to do:

1 Lower the starting sample concentration.

2 Reprime the chip. See “LabChip Kit Essential Practices” on page 23 for instructions on how to reprime the chip.

Symptom: Unexpected sharp peaks.

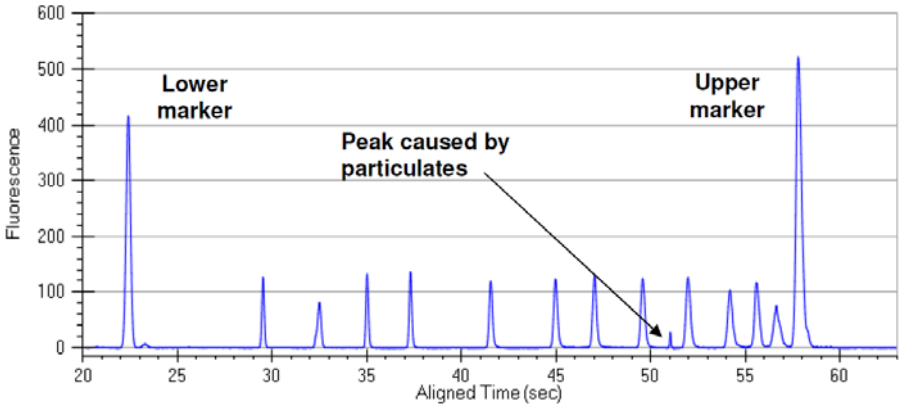

Figure 15. Unexpected sharp peak.

Possible causes:

- Dust or other particulates introduced through sample or reagents.

What to do:

1 Do one or all of the following:

- Replace the 18 megohm, 0.22-µm filtered water (Milli-Q® or equivalent) water used for chip preparation.
- Replace the buffer used for sample and reagent preparation.
- Use a 0.22-micron filter for all water and buffers used for chip, sample, and reagent preparation.
- Spin down sample plate to pellet any particulates.

Symptom: Humps in several electropherograms which do not correspond to sample data.

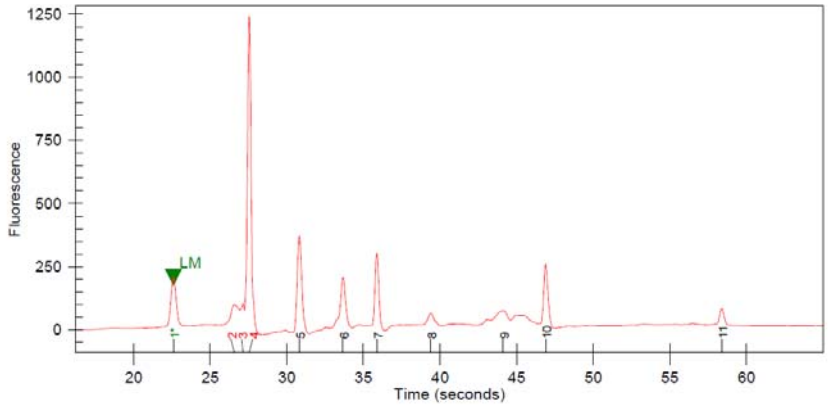

Figure 16. Humps in several electropherograms.

Possible causes:

1 Electrode 7 is dirty and has contaminated the Gel-Dye mixture in well 7.

What to do:

1 Before restarting the run, clean electrode 7. Remove the chip and follow the electrode cleaning procedure. We recommend using the provided swab and isopropanol to manually clean electrode 7.

Symptom: Peaks migrating much faster or slower than expected.

Note: *Some migration time variance between chips or within a plate is considered normal chip performance. All chips are QC tested at PerkinElmer prior to shipment.*

Possible causes:

1 Incorrect Gel-Dye ratio. Migration time is sensitive to dye concentration and peaks will migrate too fast or too slow if the dye concentration in the gel is too low or too high, respectively.

Note: *Excess dye in the separation channel will slow down migration, and less dye in the separation channel will make peaks migrate faster.*

2 Particulates from the samples may be clogging the separation channel (this will slow down migration).

3 Gel-Dye was not primed properly into the chip.

What to do:

1 Prepare a fresh Gel-Dye solution. Wash and reprime the chip with the new Gel-Dye mixture. See “LabChip Kit Essential Practices” on page 23 for instructions on how to wash and reprime the chip.

2 If fast or slow migration is observed repeatedly on a new chip, contact technical support to arrange return of the chip to PerkinElmer. Please send a data file showing the failure along with the return request.

3 Minimize the loading of particulates in the sample by performing a centrifuge spin of the sample plate (e.g. 3000 rcf for 5 minutes) before starting a new run. The debris may be flushed out of the chip by washing and re-priming the chip. See “LabChip Kit Essential Practices” on page 23 for instructions on how to wash and reprime the chip.

4 Check the O-rings on the top surface of the chip interface and clean if necessary.

LabChip Kit Essential Practices

To ensure proper assay performance, please follow the important handling practices described below.Failure to observe these guidelines may void the LabChip Kit product warranty.[1]

Note: *It is important to keep particulates out of the chip wells, channels and capillary. Many of the following guidelines are designed to keep the chips particulate-free.*

For assay and instrument troubleshooting, refer to the LabChip GX Touch software Help file or call PerkinElmer Technical Support at 1-800-762-4000.

General

- Allow the chip, sample plate, and all reagents to equilibrate to room temperature for at least 30 minutes before use.
- Clean the O-rings in the chip interface weekly and the electrodes daily. Refer to the Instrument Users Guide Maintenance and Service section for procedures.
- Avoid use of powdered gloves. Use only non-powdered gloves when handling chips, reagents, sample plates, and when cleaning the instrument electrodes and electrode block.
- Calibrate laboratory pipettes regularly to ensure proper reagent dispensing.
- Only the PerkinElmer-supplied clean room cloth can be used on the chip to clean the detection window.
- Water used for chip preparation procedures must be 18 megohm, 0.22-µm filtered water (Milli-Q® or equivalent).
- Using the “Reverse Pipetting Technique” (described next) will help avoid introducing bubbles into the chip when pipetting the gel.

1. PerkinElmer warrants that the LabChip Kit meets specification at the time of shipment, and is free from defects in material and workmanship. LabChip Kits are warranted for 90 days from the date of shipment. All claims under this warranty must be made within thirty days of the discovery of the defect.

Reverse Pipetting Technique

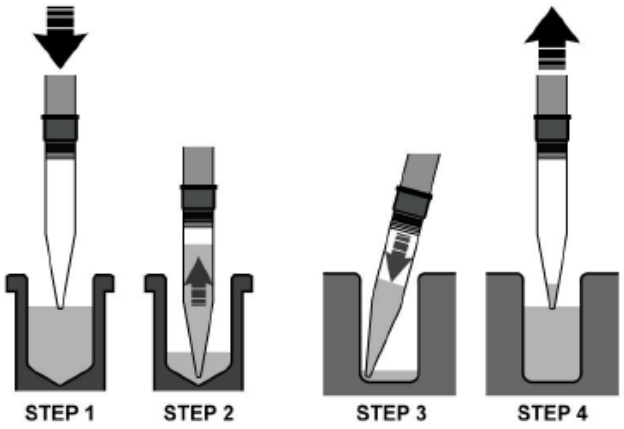

Figure 17. Reverse pipetting.

1. Depress the pipette plunger to the second stop.
2. Aspirate the selected volume plus an excess amount from the tube.
3. Dispense the selected volume into the corner of the well by depressing plunger to the first stop.
4. Withdraw the pipette from the well.

Reagents

- Store reagents at 2-8°C when not in use.
- The LabChip dye contains DMSO and should be thawed completely before use. It is recommended that you prepare aliquots to reduce the time required for thawing.
- Gently vortex all kit reagents before use.
- Dispense reagents into chip wells slowly without introducing air bubbles. Insert the pipette tip vertically and to the bottom of the chip well.
- Protect the dye and Gel-Dye mixture from light. Store in the dark at 2-8°C when not in use.
- The Gel-Dye mixture expires 3 weeks after preparation.

Chips

Repriming Chips

Note: *Buffer tubes filled with 1X DNA sample buffer or water should be placed into the instrument while priming or washing chips.*

- Touch the *Unload Chip* button on the *Home* screen to open the instrument door. Place the chip into the instrument.
- Close the chip door securely and choose the corresponding assay.
- Touch the *Prime* button on the *Home* screen to reprime the chip.

Washing and Repriming Chips

- Touch the *Unload Chip* button on the *Home* screen to open the instrument door.
- Place the chip into the plastic storage container. The sipper should be submerged in the fluid reservoir.
- Remove the reagents from each well of the chip using vacuum.
- Each active well (1, 3, 4, 7, 8, and 10) should be rinsed and aspirated twice with water (Milli-Q® or equivalent). Do not allow active wells to remain dry.
- Add 100 µL of Storage Buffer to active wells.
- Place the chip in the LabChip GX Touch/GXII Touch instrument. Ensure that a Buffer Tube with 750 µL of water (Milli-Q®or equivalent) is in the buffer slot
- Close the chip door securely.
- Touch the *Wash* button on the *Home* screen. The *Wash* screen opens (Figure 18).

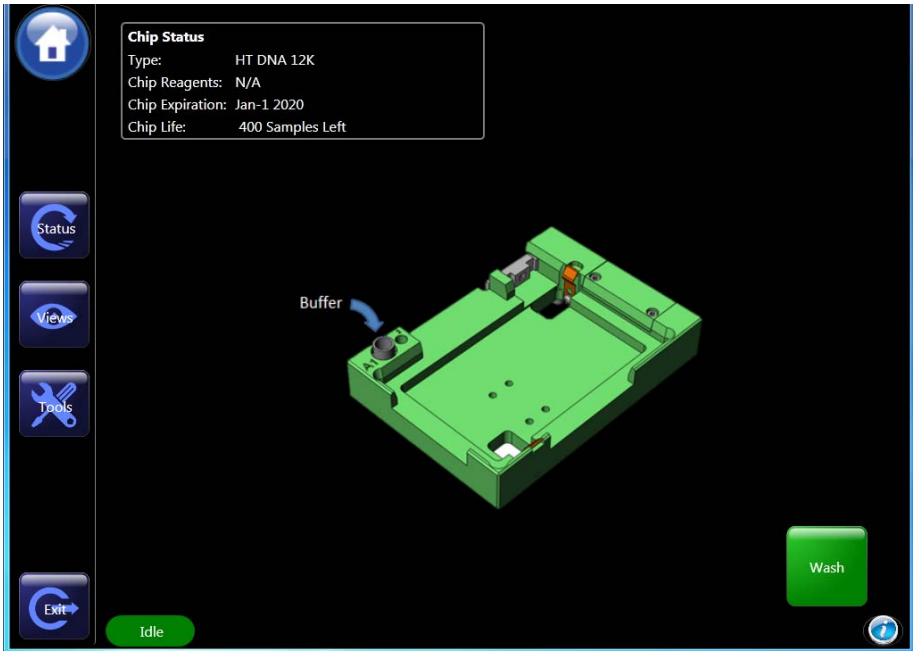

Figure 18. *Wash* screen.

- After the completion of the wash cycle return the chip to the chip container ensuring the sipper is immersed in fluid.
- Thoroughly aspirate all fluid from the chip wells using a vacuum line.
- Ensure that each active well (1, 3, 4, 7, 8, and 10) is rinsed and aspirated twice with water (Milli-Q® or equivalent). Do not allow active wells to remain dry.
- Add Gel-Dye solution to chip wells 3, 7, 8 and 10 using a Reverse Pipetting Technique as shown in Figure 17.
- Add Marker (green cap ●) to chip well 4. Use 50 µL for 96-well or 120 µL for 384- well plate analysis. Please note that the marker well may need to be replenished if the chip is in idle mode on the instrument for an extended period of time.
- Place the chip into the LabChip GX Touch/GXII Touch instrument.
- Close the chip door securely.
- Touch the *Run* or *Prime* button on the *Home* screen.

- If air bubbles are not dislodged after a reprime, apply a vacuum to the sipper. Perform this by filling all active wells with 100 µL of Chip Storage Buffer. Then suction the sipper with a vacuum line as shown in Figure 19 until droplets of fluid flow out from the sipper. When suctioning the sipper, be careful not to bend or break the sipper. To facilitate this, cut the end of the pipette tip attached to the vacuum line to widen the mouth.

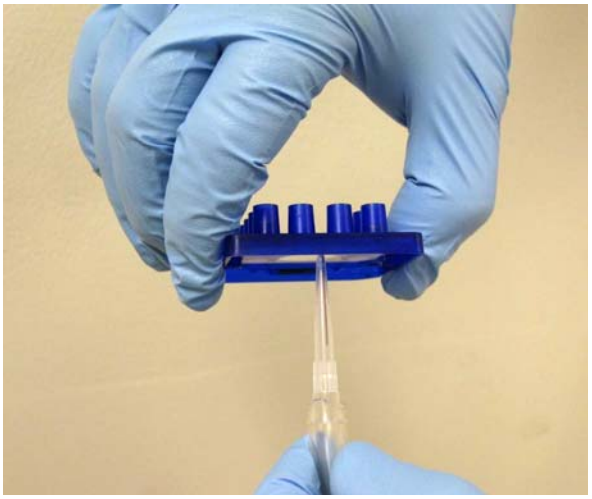

Figure 19. Removing an air bubble or clog by suctioning the sipper with a vacuum line.

Other Considerations:

- DNA chips should be stored refrigerated prior to first use.
- Cover the active wells on the chip with Parafilm® to prevent evaporation and store at 2-8°C until next use. If using the chip again within 24 hours it may be left at room temperature. Storage of chip with dry wells may cause it to become clogged.
- Do not allow the liquid in the chip container to freeze, as this may lead to poor chip performance. Do not submerge the chip in any solution.
- The entire chip surface must be thoroughly dry before use.
- The sipper must be kept immersed in fluid at all times and should not be exposed to an open environment for long periods of time.
- Use care in chip handling to prevent sipper damage. Damage to the sipper can result in inconsistent sampling.
- Avoid exposing the chips to dust by keeping them in a closed environment such as in the chip container or in the instrument before and after chip preparation.

- Chips can be prepared and left idle on the instrument for up to 8 hours. This workflow allows analysis of samples as needed throughout the day without having to re-prep the chip as long as the maximum number of samples per chip prep is not exceeded.
- PerkinElmer recommends the chip be re-prepared after it has been idle for 8 hours.

Samples

- Prepared sample plates should be free of gas bubbles and particulate debris, both of which may inhibit sipper flow.
- Sample plates containing gas bubbles and/or particulate debris should be spun down at 3000 rpm (1250 rcf) prior to analysis.
- Up to 96 samples in a 96-well or 384-well plate can be run with a single chip preparation.

Chip Well Aspiration Using a Vacuum

Aspirating with a pipette can leave used reagents in the chip wells. For this reason, PerkinElmer recommends vacuuming the wells instead. This can be accomplished by attaching a permanent pipette tip to a house vacuum line with trap (Figure 20). To avoid contamination, use a new disposable pipette tip over the permanent tip for each chip aspirated (Figure 21).

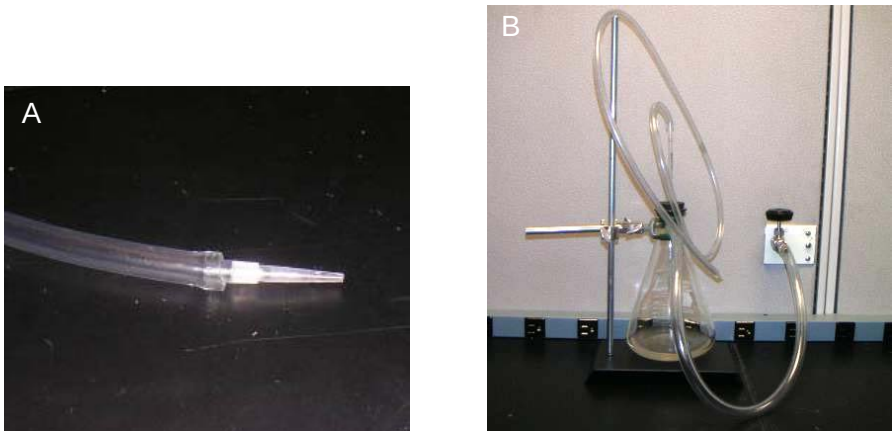

Figure 20. A: Permanent pipette tip attached to a house vacuum line; B: vacuum line with trap.

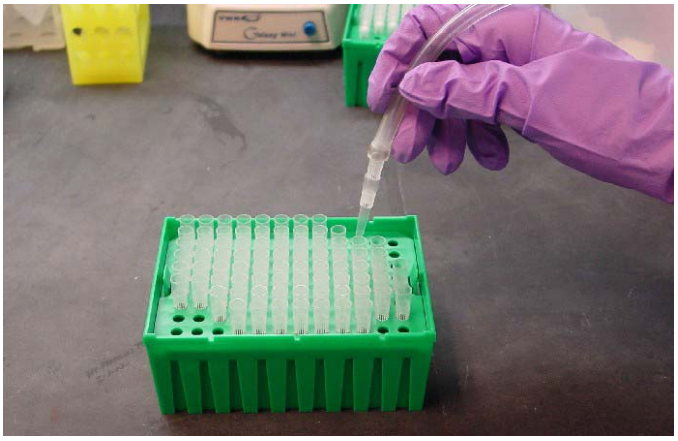

Figure 21. Replacing the disposable pipette tip.

Customer Technical Support

PerkinElmer, Inc.
68 Elm Street
Hopkinton, MA 01748-1668

PerkinElmer Technical Support
Phone (USA Toll Free): 800-762-4000
Phone (Worldwide): +1 203-925-4602
Fax: +1 203-925-4602
Email: dxsupportamericas@perkinelmer.com
Internet: www.perkinelmer.com

For additional assay and instrument troubleshooting, refer to the LabChip GX Touch Software Help file.

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http://www.perkinelmer.com

PerkinElmer, Inc.
68 Elm Street
Hopkinton, Massachusetts 01748 U.S.A.
TEL 508-435-9500
FAX 508-435-3439
http://www.perkinelmer.com

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