

RNA Assay User Guide

For LabChip® GX Touch/GXII Touch



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Specifications

Assay Specifications

Table 1. Assay Specifications

Linear Range	25 - 250 ng/μL (Std Sens) 5 - 50 ng/μL (High Sens)
Quantitation Reproducibility	20% CV
Size Range	100 - 6000 nucleotides (suitable for total RNA)
Minimum RNA Sample Volume	2 μL of user sample (Std Sens) 6 μL of user sample (High Sens)
Run Time	80 seconds per sample (about 2.5 hours for 96 samples)
Setup Time	Approximately 30 minutes to prepare chip and samples
Samples per Chip Prep	Up to 192 samples per HT chip prep Up to 48 samples per LT chip prep
Chip Preps per Reagent Kit	5 HT chip preps or 10 LT chip preps

Sample Conditions

Table 2. Sample Conditions

Additives	PerkinElmer recommends that BSA and detergents exceeding 0.05 mg/mL and 0.01% (v/v) respectively in concentration not be used. Higher concentrations can result in chip failure. In addition, non-aqueous solvents are not compatible with LabChip protocols.
Particulates	Spin down all sample plates prior to analysis. Filter all buffers with a 0.22 μm cellulose acetate filter.
Salt Concentration	Total salt concentration must not exceed 125 mM. Higher salt concentrations can alter performance and reduce assay sensitivity.







RNA Reagent Kit Contents

The RNA Reagent Kit (P/N CLS960010) contains the reagents and consumables listed in the tables below.

Note: Use only consumables that are within their expiration date.

Storage: When not in use, store reagents at the temperatures specified in [Table 3](#).

Table 3. Reagents

Reagent	Vial	Quantity	Storage Temperature
RNA Dye Concentrate	Blue 	1 vial, 0.5 mL	2-8°C
RNA Chip Storage Buffer	White 	5 vials, 1.8 mL each	2-8°C
RNA Gel Matrix	Red 	5 vials, 0.510 mL each	2-8°C
RNA Marker	Green 	1 vial, 0.8 mL	2-8°C
10X RNA Sample Buffer Concentrate	Purple 	6 vials, 2.0 mL each	2-8°C
RNA Ladder (packaged separately, P/N 760634) ^a	Yellow 	2 vials, 0.03 mL each	-70°C

a. Additional RNA Ladder can be ordered separately using Part Number 760634.

Table 4. Consumable Items

Item	Supplier and Catalog Number	Quantity
Spin Filters	Costar [®] , Cat. # 8160	6
Detection Window Cleaning Cloth	VWR [®] , Cat. # 21912-046	1
Swab	ITW Texwipe [®] , Cat. # TX758B	3
Centrifuge Tubes, 2.0 mL	(Not sold separately)	5
Ladder Tubes, 0.2 mL	(Not sold separately)	10
Buffer Tubes, 0.75 mL	(Not sold separately)	10

DNA 5K/RNA/CZE LabChips

Storage: Store chips at 2-8°C. If using a prepared chip again within 24 hours, the chip can be stored at room temperature.

Table 5. RNA LabChips

Item	Part Number	Samples per Chip
DNA 5K/RNA/CZE HT Chip (GX/GXII Touch HT)	760435	2000
DNA 5K/RNA/CZE 24 Chip (GX/GXII Touch HT or 24)	CLS138949	750

Safety and Usage

Safety Warnings and Precautions

CAUTION

We recommend that this product and components be handled only by those who have been trained in laboratory techniques and that products are used in accordance with the principles of good laboratory practice. All chemicals should be considered potentially hazardous. When handling chemical reagents, wear suitable protective clothing such as laboratory overalls, safety glasses, and gloves. Avoid contact with skin or eyes. In case of contact with skin or eyes, wash immediately with water.

WARNING!



RNA Dye contains DMSO. S24/25: Avoid contact with skin and eyes.

Usage

The RNA Assay is for use with LabChip GX Touch/GXII Touch instruments. LabChip GX Touch/GXII Touch instruments are for research use only and not for use in diagnostic procedures.

Preparation Procedures

Additional Items Required

- 18 megohm, 0.22- μ m filtered water (Milli-Q[®] or equivalent).
- 70% isopropanol solution in DI water.
- DEPC treated water (nuclease-free)
- PCR cap strips
- Bio-Rad Hard-Shell[®] 384-well Skirted PCR Plates, Cat # HSP-38XX (recommended)
- PerkinElmer Hard-Shell thin-wall 96-well skirted PCR plate (blue), Cat # 6008870 (recommended)

Preparing the Gel-Dye Solution

Warning: *The RNA dye is light sensitive. **Do not expose the Dye or Gel-Dye solution to light for any length of time.** Keep the prepared Gel-Dye solution in the dark.*

- 1 Allow the chip and refrigerated reagents to equilibrate to room temperature for at least 30 minutes before use.

Note: *The RNA Dye Concentrate contains DMSO and **must be thawed completely** before use.*

- 2 Vortex the thawed RNA Dye Concentrate (blue cap ●) for 10 - 15 seconds before use.
- 3 Transfer **75 μ L** of **RNA Dye Concentrate** (blue cap ●) to a 2.0 mL centrifuge tube provided with the reagent kit.
- 4 Add **425 μ L** of **RNA Gel Matrix** (red cap ●) using a reverse pipetting technique (for more details see [“LabChip Kit Essential Practices” on page 30](#)).
- 5 Vortex the Gel-Dye solution until it is well mixed and spin down for a few seconds.
- 6 Transfer the Gel-Dye solution to a spin filter. Use a centrifuge tube filled with 500 μ L of water to balance the centrifuge.
- 7 Centrifuge at 9300 rcf for 10 minutes at room temperature.
- 8 Discard the filter.
- 9 Label and date the tube.
- 10 Store in the dark at 2-8°C. Use within 5 days.

Preparing the RNA Samples and RNA Ladder

- 1 Prepare 1X Sample Buffer by adding **620 μ L RNA Sample Buffer Concentrate** (purple cap ●) to **5580 μ L DEPC treated (nuclease-free) water**.

Note: This volume is enough for one full 96-well plate run (includes the samples, ladder, and buffer tube). You may adjust accordingly for partial plates. Sample Buffer is stable after dilution, but to avoid RNase contamination, Sample Buffer should be prepared fresh.

- 2 Allow the RNA Ladder (yellow cap ●) to thaw on ice. (It is recommended to aliquot the RNA Ladder to five 4- μ L aliquots for individual use after thawing the vial for the first time.)
- 3 Pipette **4 μ L** of **RNA Ladder** into the provided 0.2 mL Ladder Tube and cover.
- 4 Pipette **2 μ L** (RNA Std Sens) or **6 μ L** (RNA High Sens) of **sample** into individual wells of a microtiter plate or into RNase-free microcentrifuge tubes.
- 5 Cover samples in microtiter plates with PCR cap strips. Foil is not recommended because the adhesive may contaminate the samples. If diluting the samples, use DEPC treated (nuclease-free) water.
- 6 Heat samples and ladder for 2 minutes at 70°C.
- 7 Snap cool the ladder and samples by immediately placing the tube and plate on ice for 5 minutes.
- 8 Add **46 μ L** (RNA Std Sens) or **19 μ L** (RNA High Sens) of prepared **1X Sample Buffer** (prepared in Step 1) to each sample. Mix by pipetting up and down a few times. Avoid creating air bubbles.
- 9 Cover with PCR cap strips and spin down the plate at 3000 rpm for 5 minutes at room temperature to remove air bubbles.
- 10 Add **96 μ L** of prepared **1X Sample Buffer** to the RNA Ladder. Ensure there are no air bubbles in the Ladder Tube.
- 11 Insert the Ladder Tube into the ladder slot on the LabChip GX Touch/GXII Touch instrument (see [Figure 1 on page 9](#)).

Preparing the Buffer Tube

- 1 Add **750 μ L** of prepared **1X Sample Buffer** to the 0.75 mL Buffer Tube provided with the reagent kit. Ensure there are no air bubbles in the Buffer Tube.
- 2 Insert the Buffer Tube into the buffer slot on the LabChip GX Touch/GXII Touch instrument (see [Figure 1](#)).

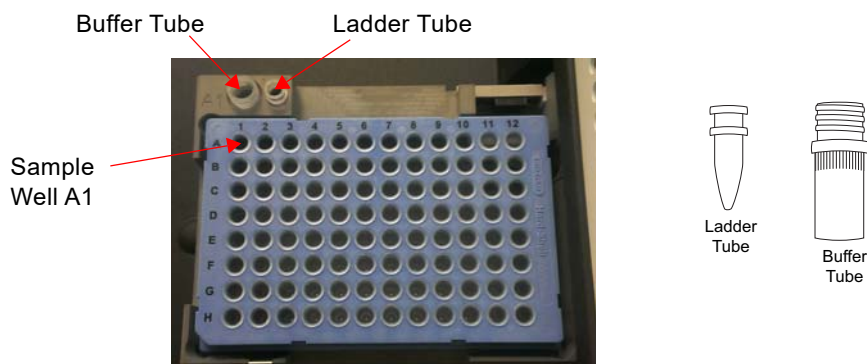


Figure 1. Locations of the Buffer Tube and Ladder Tube in the GX Touch/GXII Touch instrument

Preparing the Chip

- 1 Allow the chip to equilibrate to room temperature for at least 30 minutes before use.
- 2 Use a pipette tip attached to a vacuum line to thoroughly aspirate all fluid from the chip wells (see [Figure 2](#)). For details on how to set up a vacuum line, see [page 33](#).

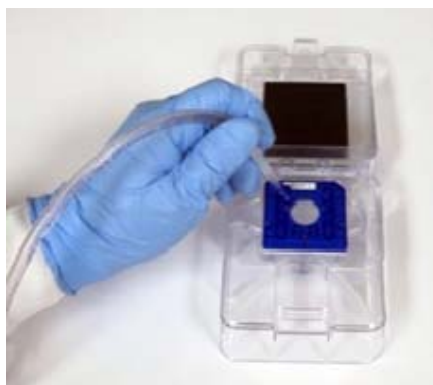


Figure 2. Using a vacuum to aspirate the chip wells is more effective than using a pipette

- 3 Rinse and completely aspirate each active chip well (1, 3, 4, 7, 8, and 10) twice with water (Milli-Q[®] or equivalent). Do not allow active wells to remain dry.
- 4 If any water spills onto the top or bottom chip surfaces during rinsing, aspirate using the vacuum line. DO NOT move the tip over the detection window. Use the provided Detection Window Cleaning Cloth dampened with water (Milli-Q[®] or equivalent) or alcohol to clean the detection window as needed.
- 5 Using a reverse pipetting technique, add **50 μ L** (Low-throughput) or **75 μ L** (High-throughput) **Gel-Dye solution** to chip wells **3, 7, 8, and 10** as shown in [Figure 3](#) or [Figure 4](#).

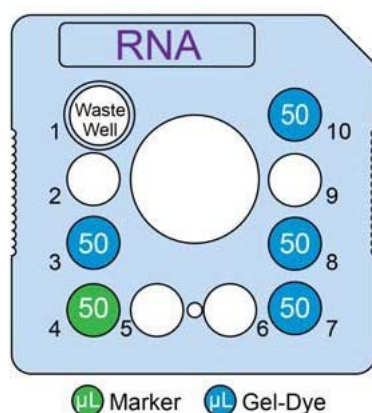


Figure 3. Low-throughput Chip Preparation

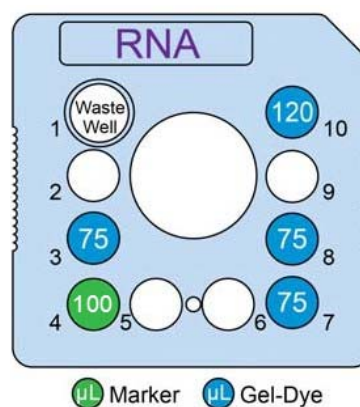


Figure 4. High-throughput Chip Preparation

- 6 Add **50 μ L** (Low-throughput) or **100 μ L** (High-throughput) **RNA Marker** (green cap ●) to chip well 4 as shown in [Figure 3](#) or [Figure 4](#).

Note: The marker well may need to be replenished if the chip is in idle mode on the instrument for an extended period of time.

- 7 Make sure the rims of the chip wells are clean and dry.
- 8 **IMPORTANT:** Ensure **chip well 1** (waste well) is empty before placing the chip into the instrument.

Inserting a Chip into the LabChip GX Touch/GXII Touch Instrument

- 1 Remove the PCR cap strips and place the sample plate into the instrument.
- 2 Check that the Buffer Tube and Ladder Tube are placed in the instrument properly.
- 3 Remove the chip from the chip storage container and inspect the detection window. Clean BOTH sides of the detection window with the PerkinElmer-supplied clean-room cloth dampened with a 70% isopropanol solution in DI water.
- 4 Touch the **Unload Chip** button on the **Home** screen ([Figure 5](#)).

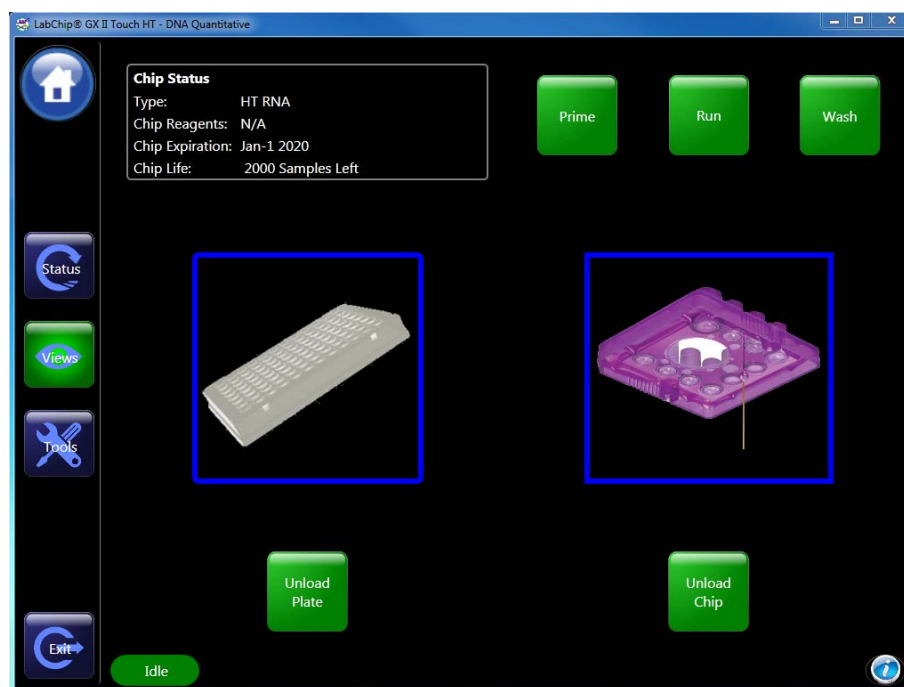


Figure 5. Home screen

- 5 Insert the chip into the LabChip GX Touch/GXII Touch instrument ([Figure 6](#)) and close the chip door securely.

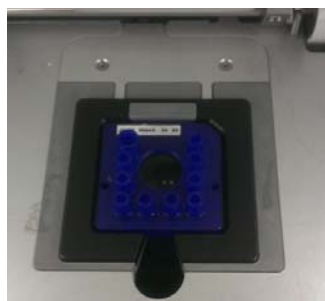


Figure 6. Chip in the LabChip GX Touch/GXII Touch instrument

- 6** Touch the **Load Plate** button on the Home screen (Figure 5) to retract the sample plate and move the sipper to the Buffer Tube. The Assay Choice screen opens (Figure 7).

Note: Do not keep the chip door open for any length of time. Dye is sensitive to light and can be photobleached.

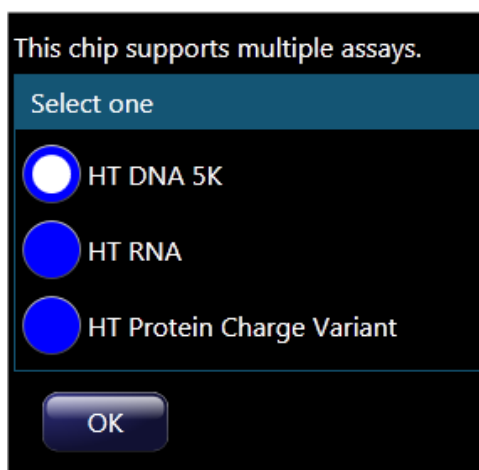


Figure 7. Assay Choice screen

- 7** Touch the desired assay and then touch **OK**.

Notes: If performing multiple chip preps in one day, wash the chip between chip preparations using the instrument and RNA Chip Storage Buffer as described in [“Cleaning and Storing the Chip” on page 20](#).

Be sure to periodically clean the O-rings on the top plate of the chip interface on the LabChip GX Touch/GXII Touch. Use the provided lint-free swab dampened with water to clean the O-rings using a circular motion. Allow the O-rings to dry before inserting a chip.

Operating Procedures

Running the Assay

Note: The chip can be primed independently from running assays on the LabChip GX Touch/GXII Touch instrument. Touch the Prime button on the Home screen (Figure 5 on page 11). Select the desired assay from the Assay drop-down list (see Figure 9). Make sure the Buffer Tube is placed in the instrument. Touch the Prime button on the Chip Priming screen.

To run an assay:

- 1 Touch the **Run** button on the Home screen (see Figure 5 on page 11). The Select Wells tab opens (see Figure 8).

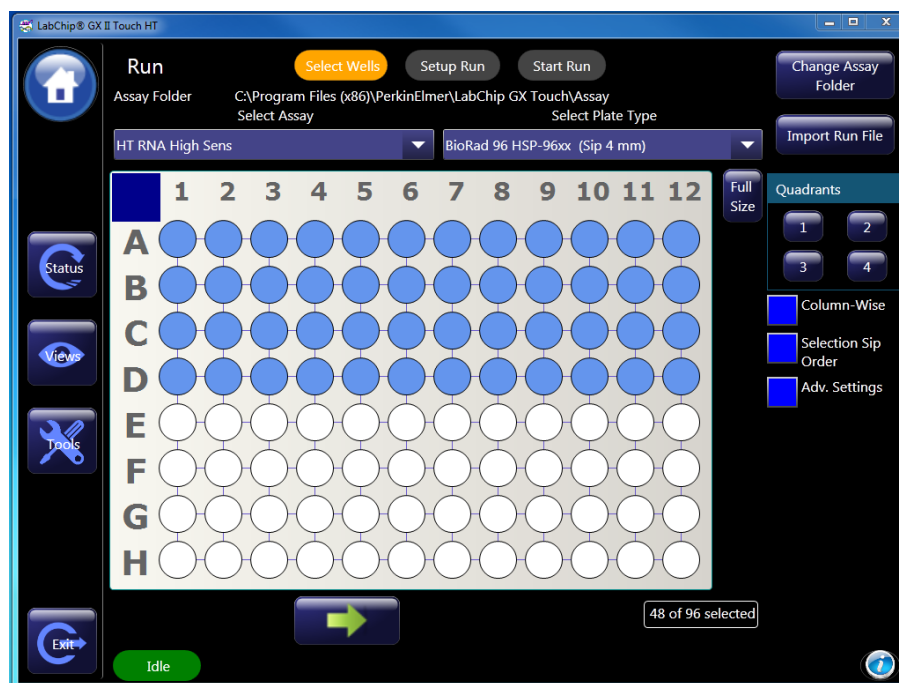


Figure 8. Select Wells Tab

- 2 Select the desired assay type (see Figure 9). For RNA assays, the valid assay types are:
 - **RNA Std Sens:** For detection of RNA in the 25 - 250 ng/μL range.
 - **RNA High Sens:** For detection of RNA in the 5 - 50 ng/μL range (requires a higher sample volume).

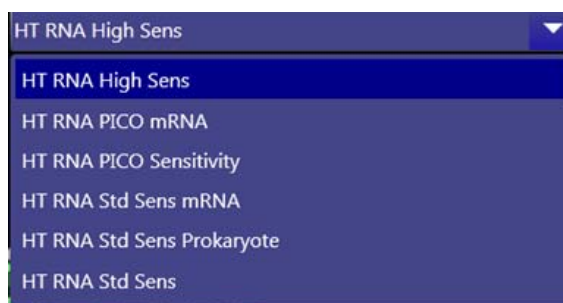


Figure 9. Assay Type Drop-Down List

- 3 Select the plate type, well pattern, and sip order (column or row). If necessary, click **Adv. Settings** to specify the number of times each well is sampled.
- 4 Touch the **Green Arrow** button. The Setup Run tab (Figure 10) opens.

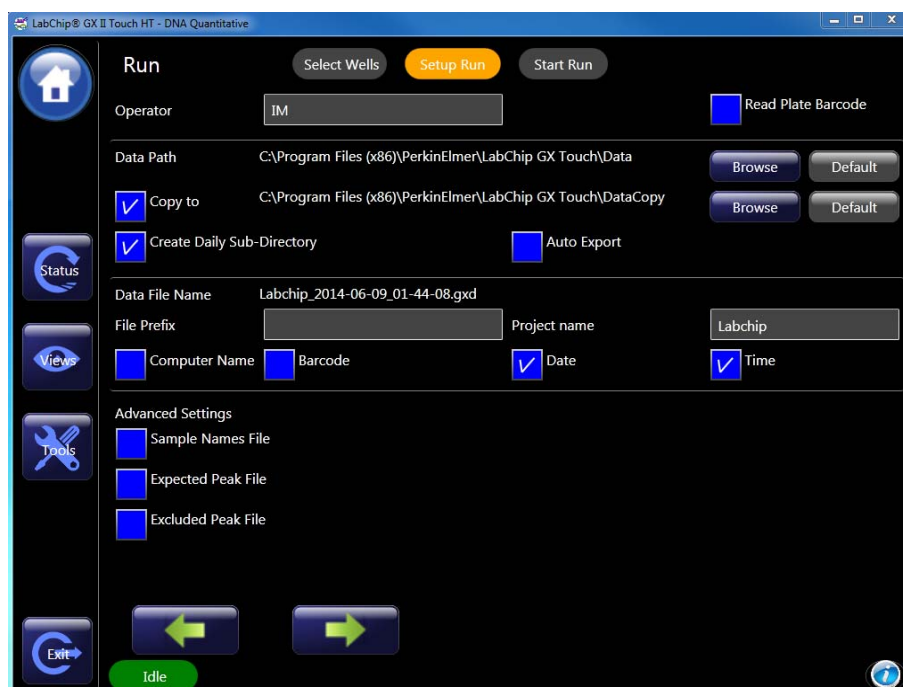


Figure 10. Setup Run Tab

- 5 Specify the operator name, the option to read the barcode, the destination of the file, the use of sample names, expected peaks, excluded peaks, the filename convention, and auto export option.
- 6 Touch the **Green Arrow** button. The Start Run tab (Figure 11) opens.



Figure 11. Start Run Tab

- 7 Touch **Start** to begin the run. If the chip has not been primed since the last time the chip door was opened, the chip is primed automatically at the start of the run.

Refreshing the Gel-Dye Solution

After running 96 samples with an HT chip prep or if the Gel-Dye solution has been in the chip for more than 3 hours (LT or HT chip prep), refresh the Gel-Dye solution in well 10.

To refresh the Gel-Dye solution:

- 1 Touch the **Unload Chip** button on the Home screen to open the instrument door.
- 2 Remove the residual Gel-Dye solution in **well 10**, either by pipette or by vacuum aspiration. The well does not need to be washed or rinsed.
- 3 Add **50 μL** (LT) or **120 μL** (HT) of fresh, unused Gel-Dye solution into **well 10**. (This could be the leftover Gel-Dye solution from an earlier filtered Gel-Dye prep). Only well 10 needs to be refreshed; you can leave the Gel-Dye solution and Marker in the other chip wells.

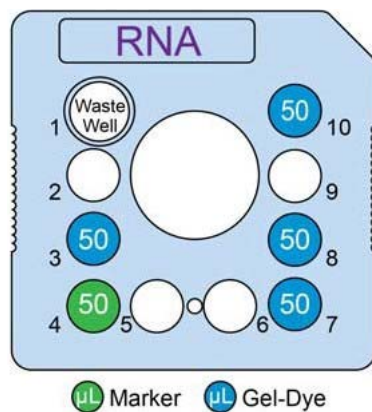


Figure 12. Low-throughput chip preparation

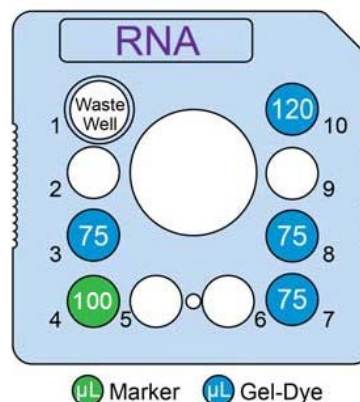


Figure 13. High-throughput chip preparation

- 4 Close the instrument door securely.
- 5 Start the next run.

Repriming Chips

If air bubbles or clogs in the chip channels are suspected, the chip can be reprimed to help remove air bubbles or clogs.

Note: Place a Buffer Tube with prepared 1X Sample Buffer solution or water into the instrument while priming or washing chips.

- 1 Touch the **Unload Chip** button on the Home screen to open the instrument door. The software automatically resets to require priming prior to running the chip again.
- 2 Place the chip and Buffer Tube into the instrument.
- 3 Close the chip door securely and choose the corresponding **assay**.
- 4 Touch the **Prime** button on the Home screen. The Prime screen opens.
- 5 Touch the **Prime** button on the Prime screen to reprime the chip.

Washing and Repriming Chips

Washing the chip clears all reagents from the chip channels. The chip can be immediately reprimed to help remove air bubbles, clogs, particulates, or residue.

- 1 Touch the **Unload Chip** button on the Home screen.
- 2 Remove the chip from the instrument and place the chip to the chip container, ensuring the sipper is submerged in fluid.
- 3 Thoroughly aspirate all fluid from the chip wells using a vacuum.
- 4 Rinse and completely aspirate each active well (1, 3, 4, 7, 8, and 10) twice with water (Milli-Q[®] or equivalent). Do not allow active wells to remain dry.
- 5 Add **120 µL** of RNA Chip Storage Buffer to each active well (**1, 3, 4, 7, 8, and 10**).
- 6 Place the chip in the LabChip GX Touch/GXII Touch instrument.
- 7 Place a Buffer Tube with **750 µL** of prepared **1X Sample Buffer** or water in the instrument.
- 8 Close the chip door securely.
- 9 Touch the **Wash** button on the Home screen. The Wash screen (Figure 14) opens.

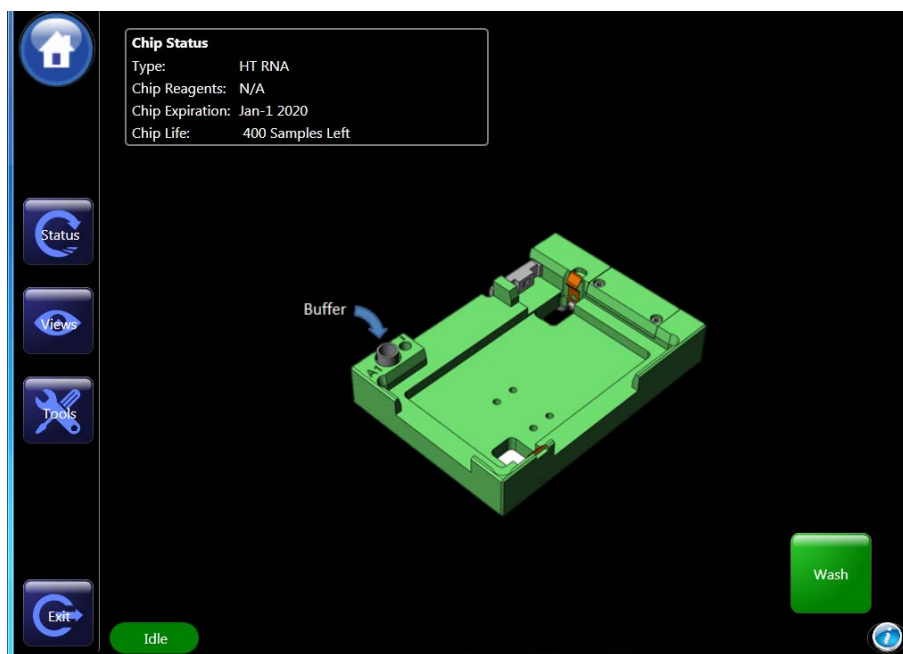


Figure 14. Wash screen

- 10 Touch the **Wash** button on the Wash screen to start the chip wash.
- 11 After the completion of the wash cycle, touch the Unload Chip button on the Home window to open the instrument door.
- 12 Return the chip to the chip container. Verify the sipper is submerged in fluid.
- 13 Thoroughly aspirate all fluid from the chip wells using a vacuum line.
- 14 Prepare the chip as described on [page 9](#).
- 15 Place the chip into the LabChip GX Touch/GXII Touch instrument.
- 16 Close the chip door securely.
- 17 Touch the **Run** or **Prime** button on the Home screen.
- 18 If air bubbles are not dislodged after a reprime, apply a vacuum to the sipper. Fill all active wells with 100 μ L of RNA Chip Storage Buffer, then suction the sipper with a vacuum line as shown in [Figure 15](#) until droplets of fluid flow out from the sipper. When suctioning the sipper, be careful not to bend or break the sipper. To facilitate this, cut the end of the pipette tip attached to the vacuum line to widen the mouth.

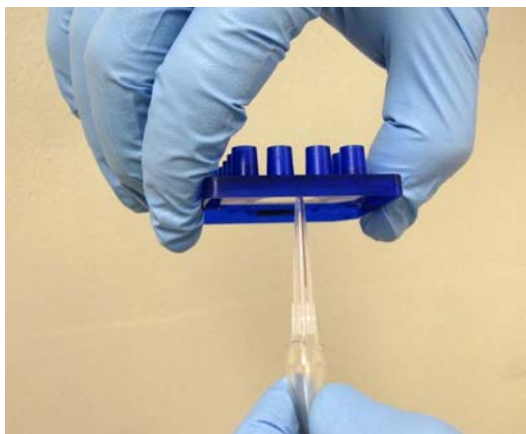


Figure 15. Removing an air bubble or clog by suctioning the sipper with a vacuum line

Cleaning and Storing the Chip

After use, the chip must be cleaned and stored in the chip container. The chip can be washed the following morning when running overnight.

- 1 Place the chip into the chip storage container. Verify the sipper is submerged in the fluid reservoir.
- 2 Remove the reagents from each well of the chip using vacuum.
- 3 Rinse and completely aspirate each active well (1, 3, 4, 7, 8, and 10) twice with water (Milli-Q[®] or equivalent).
- 4 Add **120 µL of RNA Chip Storage Buffer** (white cap ○) to the active wells.
- 5 Place the chip in the LabChip GX Touch/GXII Touch instrument.
- 6 Touch the **Wash** button in the upper right corner of the Home screen. The Wash screen opens as shown in [Figure 16](#).

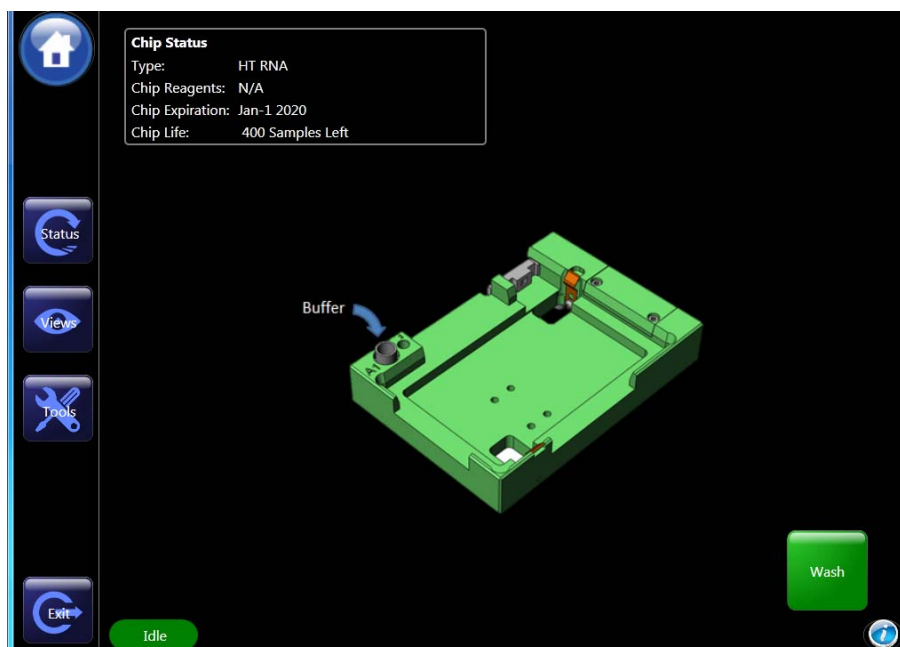


Figure 16. Wash screen

- 7 Touch the **Wash** button to start the chip wash.
- 8 When the chip wash is complete, remove the chip from the instrument and place the chip in the chip storage container.
- 9 Add an additional **50 µL RNA Chip Storage Buffer** to **well 1**.
- 10 Cover the wells with Parafilm[®] to prevent evaporation and store at 2-8°C. Storing a chip with dry wells may clog the chip. If using the chip again within 24 hours, the chip can be stored at room temperature.

Chip Cartridge Cleaning

Daily

- 1 Inspect the inside of the chip cartridge and O-rings for debris.
- 2 Use the provided lint-free swab dampened with water (Milli-Q[®] or equivalent) to clean the O-rings using a circular motion. If the O-rings stick to the chip or a pressure leak is detected, perform the more extensive monthly cleaning procedure.
- 3 Clean the electrodes with the provided lint-free swab dampened with water (Milli-Q[®] or equivalent).

Monthly

- 1 To reduce pressure leaks at the chip interface, clean the O-rings frequently. Remove the O-rings from the top plate of the chip interface on the LabChip GX Touch/GXII Touch instrument. Soak O-rings in water (Milli-Q[®] or equivalent) for a few minutes. Clean the O-ring faces by rubbing between two fingers. Wear gloves.
- 2 To reduce the occurrence of current leaks, clean the chip interface frequently. Clean the top plate of the chip interface using the provided lint free swab dampened with water (Milli-Q[®] or equivalent).
- 3 Allow the O-rings and chip interface to air dry. Reinsert the O-rings into the chip cartridge.

Results

RNA Ladder Result

The electropherogram of a typical RNA ladder is shown in [Figure 17](#).

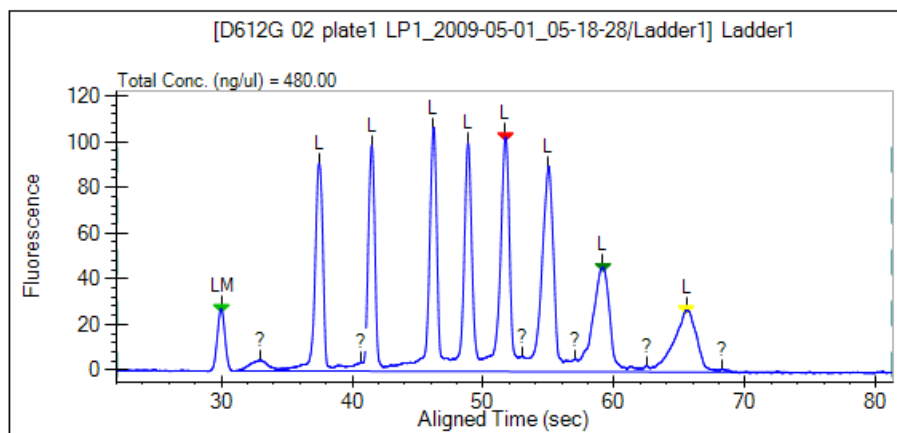


Figure 17. Typical RNA ladder

RNA Eukaryote Sample Result

The electropherogram for typical total RNA samples is shown in [Figure 18](#). Your results may vary depending on the type of total RNA.

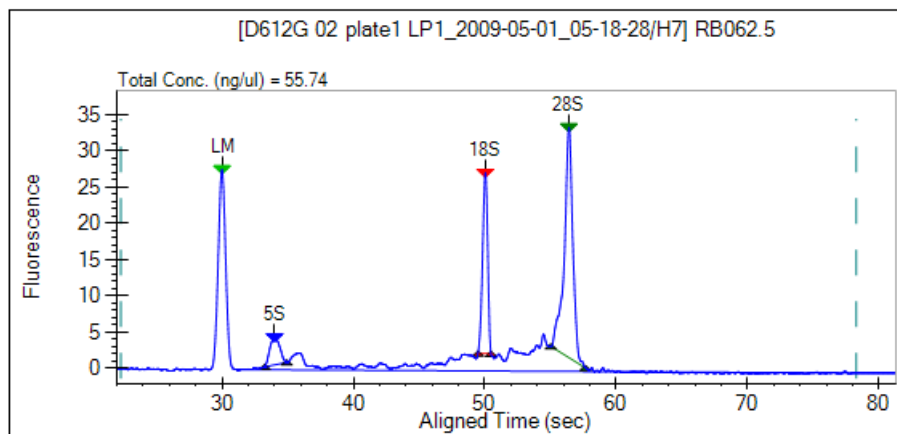


Figure 18. Electropherogram for a typical total RNA sample

RNA Prokaryote Sample Result

The electropherogram for typical Prokaryote Total RNA samples is shown in [Figure 20](#). Your results may vary depending on the type of total RNA.

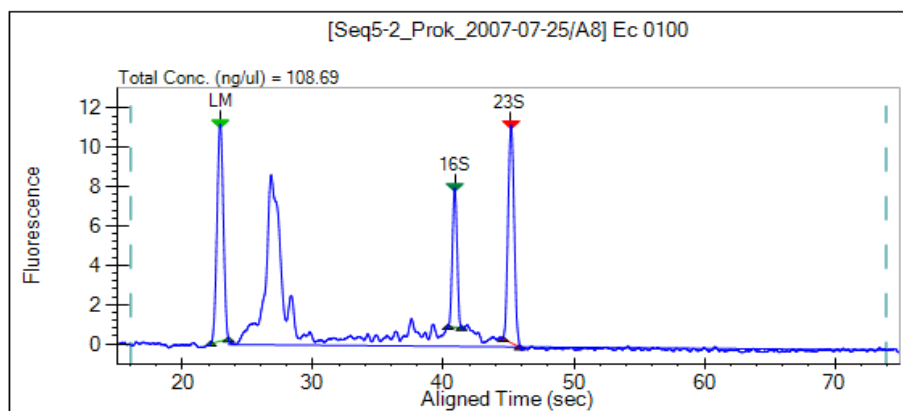


Figure 19. Electropherogram for a typical prokaryote RNA sample

RNA mRNA Sample Result

The electropherogram for a typical mRNA sample is shown in [Figure 20](#). Your results may vary depending on the type and concentration of mRNA.

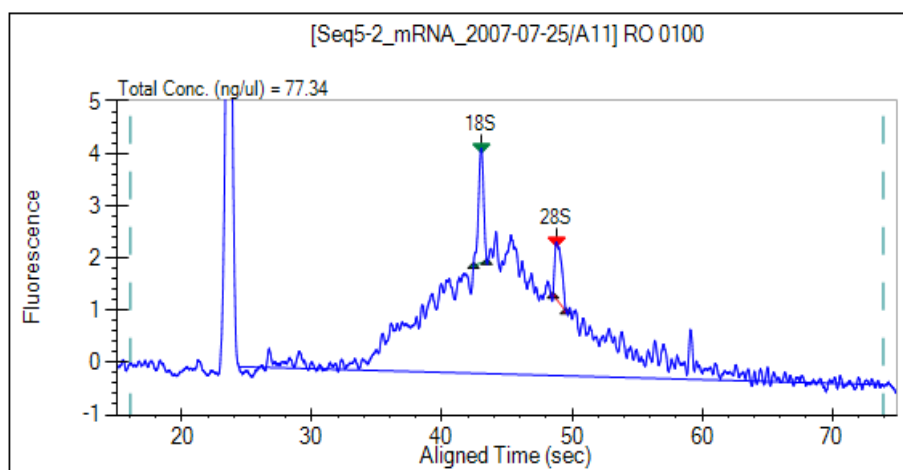


Figure 20. Electropherogram for a typical mRNA sample

Troubleshooting

***Note:** Some of the data examples shown in this section were generated with assays other than the assay described in this user guide.*

Symptom: No ladder or sample peaks but marker peaks detected.

***Note:** The lower marker peak height will most likely be greater than normal height.*

Possible causes:

- 1 Air bubble in sipper introduced during chip priming.

What to do:

- 1 Reprime the chip. See [“Repriming Chips” on page 17](#) for instructions on how to reprime the chip.

Symptom: Missing sample, ladder *and* marker peaks.

Possible causes:

- 1 Clog in sipper or marker channel of chip.

What to do:

- 1 Reprime the chip. See [“Repriming Chips” on page 17](#) for instructions on how to reprime the chip.

Symptom: Ladder detected but no sample peaks.**Possible causes:**

- 1 The sipper is not reaching the sample due to low sample volume in the well of the plate.
- 2 If the missing sample peaks occurred only in a few wells of the plate, check those wells for air bubbles.
- 3 The sipper is not reaching the sample due to an incorrect capillary height setting or incorrect plate definition.
- 4 If the plate has been uncovered for some time, sample evaporation might have occurred.
- 5 Debris from the sample or sample prep is clogging the sipper.

What to do:

- 1 Add more sample to the well.
- 2 Manually insert a larger volume pipette tip (~100 μ L) into the sample well and dislodge the bubble. Rerun these sample wells.
- 3 Check the plate definitions.
- 4 Check the sample wells, especially around the edge of the plate where evaporation is fastest, and make a fresh plate if volumes are low.
- 5 If there may be debris in the samples, spin the sample plate down in a centrifuge (e.g. 3000 rpm for 5 minutes). Unclog the sipper by repriming the chip. See ["Repriming Chips" on page 17](#) for instructions on how to reprime the chip.

Symptom: No ladder peaks but sample peaks and marker peaks are present.**Possible causes:**

- 1 Low or no ladder volume in the Ladder Tube.

What to do:

- 1 Add more ladder to the Ladder Tube and restart the run. Recommended standard ladder volume is 120 μ L (minimum volume is 100 μ L).

Symptom: No marker peaks but sample peaks are present.**Possible causes:**

- 1 No RNA Marker in chip well 4. The RNA marker may not have been put into the marker well during chip prep or the chip may have remained idle in the instrument for an extended period of time.
- 2 If there is RNA Marker in chip well 4, the marker channel may be clogged.

What to do:

- 1 Add or replenish the RNA Marker in the chip:
 - a Touch the **Unload Chip** button on the Home screen to open the chip door.
 - b Return the chip to the chip container, ensuring the sipper is immersed in fluid.
 - c Thoroughly aspirate all fluid from chip well 4 using a vacuum line.
 - d Rinse and completely aspirate chip well 4 twice with water (Milli-Q® or equivalent).
 - e Add RNA Marker (green cap ●) to chip well 4.
 - f Reinsert the chip back into the instrument.
 - g Restart the run.
- 2 Reprime the chip to unclog the marker channel. See [“Repriming Chips” on page 17](#) for instructions on how to reprime the chip.

Symptom: Ladder traces show up in the lanes following the ladders (delayed sip).

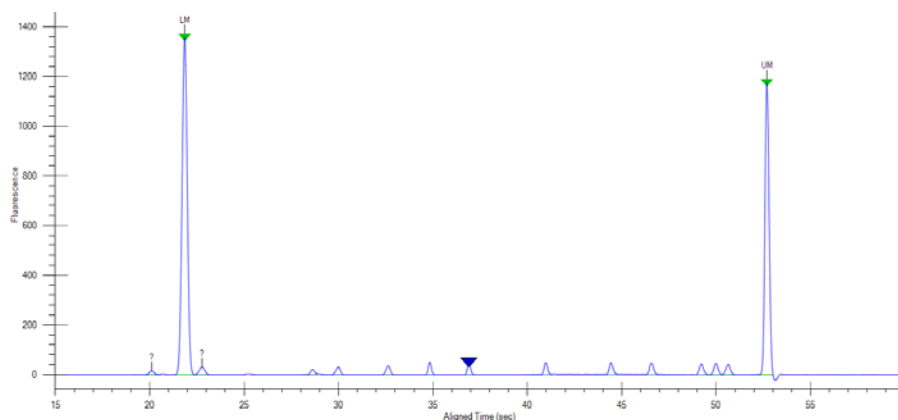


Figure 21. Small ladder peaks in sample well caused by delayed sip

Possible causes:

- 1 Separation channel overloaded with sample.
- 2 Partial clog in the separation channel.

What to do:

- 1 Lower the starting sample concentration.
- 2 Reprime the chip. See [“Repriming Chips” on page 17](#) for instructions on how to reprime the chip.

Symptom: Unexpected sharp peaks.

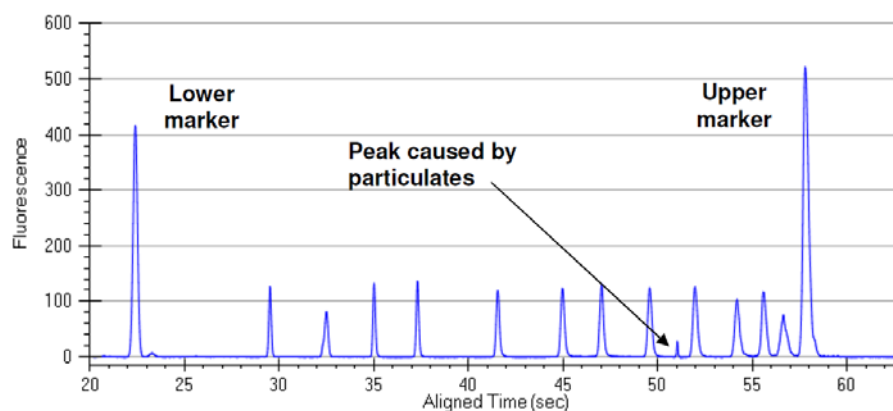


Figure 22. Unexpected sharp peak

Possible causes:

- Dust or other particulates introduced through sample or reagents.

What to do:

- 1 Do one or all of the following:
 - Replace the 18 megohm, 0.22- μ m filtered water (Milli-Q® or equivalent) used for chip preparation.
 - Replace the prepared 1X Sample Buffer used for sample and reagent preparation.
 - Use a 0.22-micron filter for all water and buffers used for chip, sample, and reagent preparation.
 - Spin down sample plate to pellet any particulates.

Symptom: Humps in several electropherograms which do not correspond to sample data.

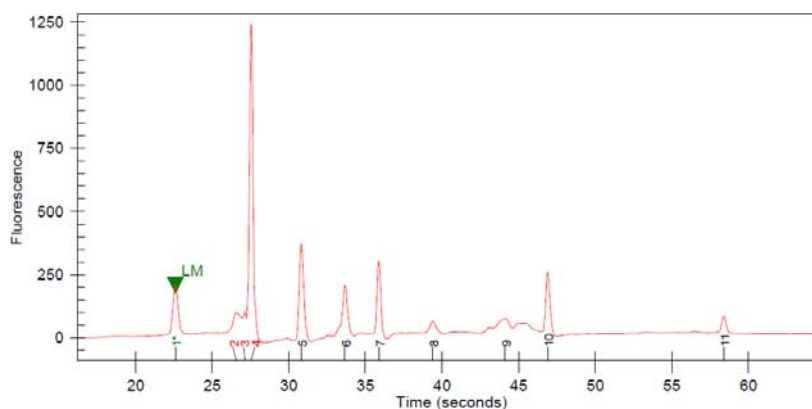


Figure 23. Humps in several electropherograms

Possible causes:

- 1 Electrode 7 is dirty and has contaminated the Gel-Dye solution in well 7.

What to do:

- 1 Before restarting the run, clean electrode 7. Remove the chip and follow the electrode cleaning procedure. We recommend using the provided swab and isopropanol to manually clean electrode 7.

Symptom: Peaks migrating much faster or slower than expected.

Note: Some migration time variance between chips or within a plate is considered normal chip performance. All chips are QC tested at PerkinElmer prior to shipment.

Normal migration time windows for the markers are:

- RNA Pico Sensitivity Assay Lower Marker: 28 - 33 seconds

Possible causes:

- 1 Incorrect Gel-Dye ratio. Migration time is sensitive to dye concentration and peaks will migrate too fast or too slow if the dye concentration in the gel is too low or too high, respectively.

Note: Excess dye within the separation channel will slow down migration, and less dye in the separation channel will make peaks migrate faster.

- 2 Particulates from the samples may be clogging the separation channel (this will slow down migration).
- 3 Gel-Dye solution was not primed properly into the chip.
- 4 A pressure leak or current leak can slow peak migration.

What to do:

- 1 Prepare a fresh Gel-Dye solution. Wash and reprime the chip with the new Gel-Dye solution. See [“Washing and Repriming Chips” on page 18](#) for instructions on how to wash and reprime the chip.
- 2 If fast or slow migration is observed repeatedly on a new chip, contact technical support to arrange return of the chip to PerkinElmer. Please send a data file showing the failure along with the return request.
- 3 Minimize the loading of particulates in the sample by spinning down the sample plate in a centrifuge (e.g. 3000 rpm for 5 minutes) before starting a new run. Flush the debris out of the chip by washing and re-priming the chip. See [“Washing and Repriming Chips” on page 18](#) for instructions on how to wash and reprime the chip.
- 4 Check the O-rings on the top surface of the chip interface and clean if necessary.

LabChip Kit Essential Practices

To ensure proper assay performance, please follow the important handling practices described below. Failure to observe these guidelines may void the LabChip Kit product warranty.¹

Note: *It is important to keep particulates out of the chip wells, channels, and capillary. Many of the following guidelines are designed to keep the chips particulate-free.*

For assay and instrument troubleshooting, refer to the LabChip GX Touch Software Help file or contact PerkinElmer Technical Support (see [page 35](#)).

General

- Allow the chip, sample plate, and all refrigerated reagents to equilibrate to room temperature for at least 30 minutes before use. Allow the RNA Ladder to thaw on ice. (It is recommended to aliquot the RNA Ladder to five 4- μ L aliquots for individual use after thawing the vial for the first time.)
- Use only consumables that are within their expiration date.
- Clean the O-rings in the chip interface weekly and the electrodes daily. Refer to the Instrument Users Guide *Maintenance and Service* section for procedures.
- Avoid use of powdered gloves. Use only non-powdered gloves when handling chips, reagents, sample plates, and when cleaning the instrument electrodes and electrode block.
- Calibrate laboratory pipettes regularly to ensure proper reagent dispensing.
- Only the PerkinElmer-supplied clean room cloth can be used on the chip to clean the detection window.
- Water used for chip preparation procedures must be 18 megohm, 0.22- μ m filtered water (Milli-Q[®] or equivalent).
- Using the Reverse Pipetting Technique on [page 31](#) will help avoid introducing bubbles into the chip when pipetting the gel.

1. PerkinElmer, Inc. warrants that the LabChip Kit meets specification at the time of shipment, and is free from defects in material and workmanship. LabChip Kits are warranted for 90 days from the date of shipment. All claims under this warranty must be made within thirty days of the discovery of the defect.

Reverse Pipetting Technique

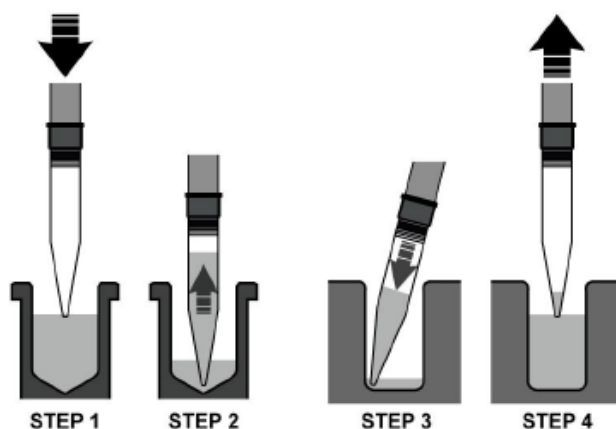


Figure 24. Reverse pipetting

- 1 Depress the pipette plunger to the second stop.
- 2 Aspirate the selected volume plus an excess amount from the tube.
- 3 Dispense the selected volume into the corner of the well by depressing the plunger to the first stop.
- 4 Withdraw the pipette from the well.

Reagents

- Store reagents at 2-8°C when not in use. Store RNA Ladder at -70°C.
- All refrigerated reagents must equilibrate to room temperature (20 - 25°C) for at least 30 minutes before use.
- Allow the RNA Ladder (yellow cap ●) to thaw on ice. (Aliquot the RNA Ladder to five 4-μL aliquots for individual use after thawing for the first time.)
- Protect the Dye and Gel-Dye solution from light. Store in the dark at 2-8°C when not in use.
- The RNA Dye contains DMSO and must be thawed completely before use. It is recommended that you prepare aliquots to reduce the time required for thawing.
- Gently vortex all kit reagents before use.
- Dispense reagents into chip wells slowly without introducing air bubbles. Insert the pipette tip vertically and to the bottom of the chip well.
- The Gel-Dye solution expires 5 days after preparation.

Chips

- Store chips at 2-8°C.
- After use, cover the active chip wells with Parafilm® and store at 2-8°C. If using the chip again within 24 hours, the chip can be stored at room temperature.
- Do not allow the liquid in the chip container to freeze, as this may lead to poor chip performance. Do not submerge the chip in any solution.
- The entire chip surface must be thoroughly dry before use.
- The sipper must be kept submerged in fluid at all times and should not be exposed to an open environment for long periods of time.
- Use care in chip handling to prevent sipper damage. Damage to the sipper can result in inconsistent sampling.
- Avoid exposing the chip to dust by keeping the chip in a closed environment such as in the chip container or in the instrument before and after chip preparation.
- Chips can be prepared and left idle on the instrument for up to 8 hours. (The Gel-Dye solution should be replaced after 3 hours.) This workflow allows analysis of samples as needed throughout the day without having to re-prepare the chip as long as the maximum number of samples per chip prep is not exceeded.
- PerkinElmer recommends the chip be re-prepared after it has been idle for 8 hours.

Samples

- Prepared sample plates should be free of gas bubbles and particulate debris, both of which may inhibit sipper flow.
- Spin down sample plates containing gas bubbles and/or particulate debris at 3000 rpm (1250 rcf) for 5 minutes prior to analysis.
- Up to 192 samples in 96-well or 384-well plates can be run with a single HT chip preparation. (See [“Refreshing the Gel-Dye Solution” on page 16](#) for instructions on refreshing the Gel-Dye solution if running more than 96 samples.) Up to 48 samples in 96-well or 384-well plates can be run with a single LT chip preparation.

Chip Well Aspiration Using a Vacuum

Aspirating with a pipette can leave used reagents in the chip wells. For this reason, PerkinElmer recommends vacuuming the wells instead. This can be accomplished by attaching a permanent pipette tip to a house vacuum line with trap (Figure 25). To avoid contamination, use a new disposable pipette tip over the permanent tip for each chip aspirated (Figure 26).

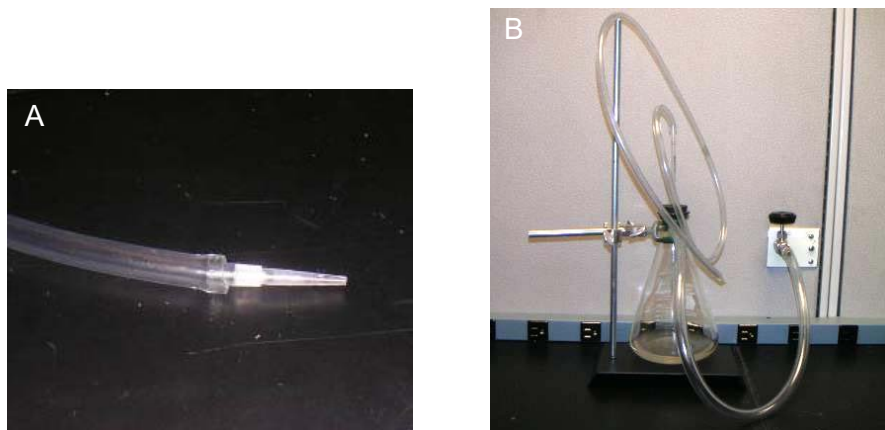


Figure 25. A: Permanent pipette tip attached to a house vacuum line; B: vacuum line with trap



Figure 26. Replacing the disposable pipette tip

Reordering Information

Table 6. Reordering Information

Product	Part Number
DNA 5K/RNA/CZE LabChip for LabChip GX Touch/GXII Touch HT	760435
DNA 5K/RNA/CZE LabChip for LabChip GX Touch/GXII Touch 24	CLS138949
RNA Reagent Kit	CLS960010
Detection Window Cleaning Cloth	VWR [®] , Cat. # 21912-046
Swab	ITW Texwipe [®] , Cat. # TX758B
Spin Filters	Costar [®] , Cat. # 8160

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Internet: www.perkinelmer.com

For additional assay and instrument troubleshooting, refer to the LabChip GX Touch Software Help file or the *LabChip GX Touch User Manual*.

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