

## **Human C-X-C Motif Chemokine 9/Monokine Induced by Gamma Interferon (CXCL9/MIG) LANCE *Ultra* Detection kit**

Product number: TRF1280

Caution: For Laboratory Use. A research product for research purposes only.

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## ○ Product Information

- Application:** This kit is designed for the quantitative determination of CXCL9/MIG in culture media using a homogeneous LANCE *Ultra* assay (no wash steps).
- Sensitivity:** Lower Detection Limit (LDL): 10.9 pg/mL  
Lower Limit of Quantification (LLOQ): 55.5 pg/mL  
EC<sub>50</sub>: 4.6 ng/mL
- Dynamic range:** Kit designed to detect CXCL9/MIG between: 10.9 – 30,000 pg/mL (Figure 1).

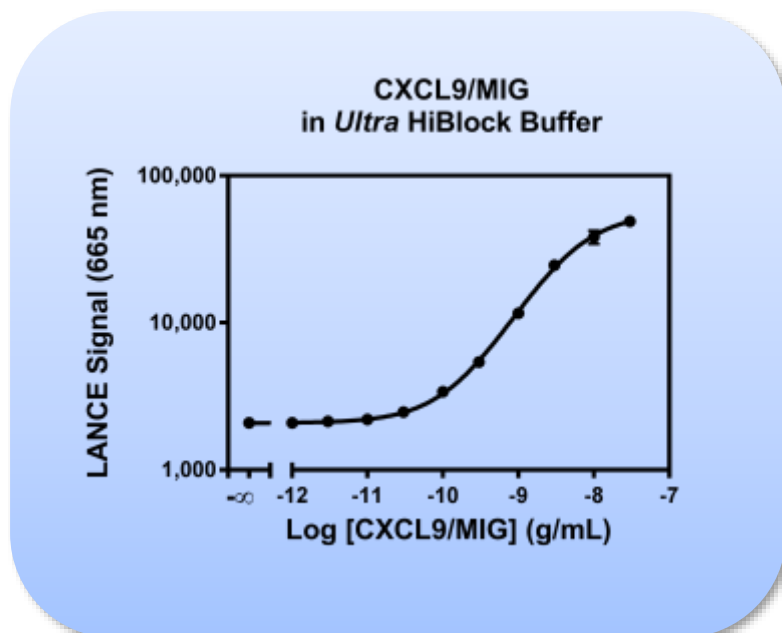


Figure 1. Typical sensitivity curves in *Ultra* HiBlock Buffer. The data was generated using a white Optiplat<sup>TM</sup>-384 microplate and the VICTOR X, ViewLux, EnVision or EnSpire Multilabel Plate Reader equipped with TR-FRET option

- Storage:** Store kit in the dark at +4°C. Store reconstituted analyte at -20°C.
- Stability:** This kit is stable for at least 12 months from the manufacturing date when stored in its original packaging and the recommended storage conditions. Note: Once reconstituted, the CXCL9/MIG analyte is stable for at least 75 days when stored at -20°C.

## ○ Quality Control

Lot to lot consistency is confirmed in an LANCE *Ultra* assay. EC<sub>50</sub> and LDL were measured on the VICTOR X, ViewLux, EnVision or EnSpire Multilabel Plate Reader equipped with TR-FRET option using the protocol described in this technical data sheet. We certify that these results meet our quality release criteria. Maximum counts may vary between lots and the instrument used, with no impact on LDL measurement.

## ○ Analyte of Interest

C-X-C Motif Chemokine 9 (CXCL9), previously called MIG, is a 14 kDa protein belonging to the intercrine alpha (chemokine CXC) family. Its induction is enhanced by TNF $\alpha$  in dermal fibroblasts and vein endothelial cells. The synthesis of CXCL9 is specifically induced in macrophages, monocytes, neutrophils, APC, B cells, and eosinophils by IFN $\gamma$  and mediated via the JAK-STAT signaling pathway. The main function of this chemokine is the recruitment of leukocytes to sites of infection and inflammation. Some studies have shown that CXCL9 is active against Gram-negative and Gram-positive bacteria. CXCL9 may play a role as a mediator of T-cell recruitment and activation in some diseases like psoriasis and pulmonary disease. CXCL9 is expressed in allogeneic skin grafts several days before completion of rejection.

## ○ Description of the LANCE *Ultra* Assay

LANCE® and LANCE® (Lanthanide chelate excite) *Ultra* are our TR-FRET (time-resolved fluorescence resonance energy transfer), homogeneous (no wash) technologies. One antibody of interest is labeled with a donor fluorophore (a LANCE Europium chelate) and the second molecule is labeled with an acceptor fluorophore [ULight™ dye]. Upon excitation at 320 or 340 nm, energy can be transferred from the donor Europium chelate to the acceptor fluorophore if sufficiently close for FRET (~10 nm). This results in the emission of light at 665 nm.

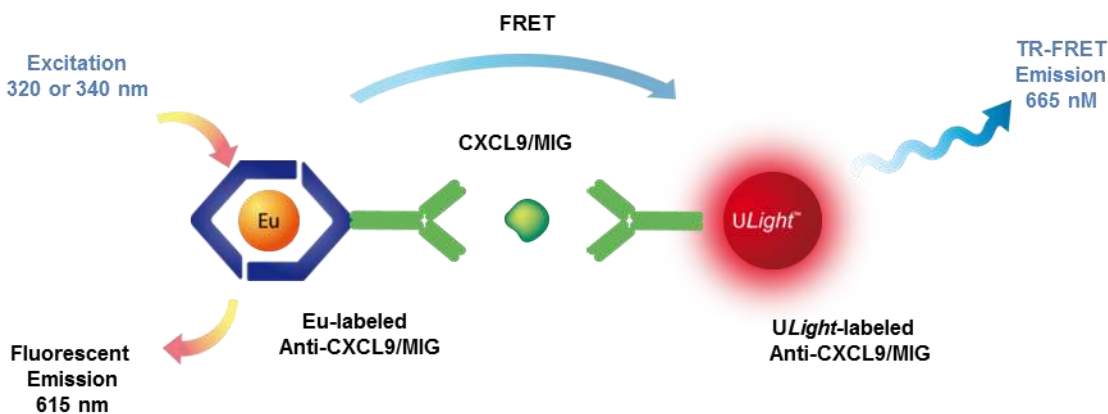


Figure 2. LANCE assay principle.

○ **Precautions**

- All blood components and biological materials should be handled as potentially hazardous.
- Some analytes are present in saliva. Take precautionary measures to avoid contamination of the reagent solutions.

○ **Kit Content: Reagents and Materials**

Kit components	TRF1280C (500 assay points <sup>***</sup> )	TRF1280M (10 000 assay points <sup>***</sup> )
LANCE <i>Ultra</i> Eu-labeled Anti-CXCL9/MIG Antibody stored TSA, 0.1% BSA	10 µL @ 500 nM (1 clear tube, yellow cap)	120 µL @ 500 nM (1 clear tube, orange cap)
LANCE <i>Ultra ULight</i> -labeled Anti-CXCL9/MIG Antibody stored in TSA, 0.1% BSA	60 µL @ 500 nM (1 brown tube, blue cap)	1200 µL @ 500 nM (1 brown tube, green cap)
CXCL9/MIG Analyte * lyophilized	0.3 µg (1 tube, <u>clear</u> cap)	0.3 µg (1 tube, <u>clear</u> cap)
<i>Ultra</i> HiBlock Buffer (5X) **	2 mL, 1 small bottle	100 mL, 1 large bottle

\* Reconstitute CXCL9/MIG in 100 µL Milli-Q® grade H<sub>2</sub>O. The reconstituted analyte should be used within 60 minutes or aliquoted into screw-capped polypropylene vials and stored at -20°C for further experiments. Avoid multiple freeze-thaw cycles. It has been demonstrated that reconstituted CXCL9/MIG is stable for at least 75 days at -20°C. One vial contains an amount of CXCL9/MIG sufficient for performing 10 standard curves. Additional vials can be ordered separately (cat # TRF1280S).

\*\* Extra buffer can be ordered separately (cat # TRF1011C: 10 mL, cat # TRF1011F: 100 mL). 5X *Ultra* HiBlock Buffer may appear cloudy, especially after storage at cold temperature. Agitate and/or stir at room temperature to redissolve prior to dilution.

\*\*\* The number of assay points is based on an assay volume of 20 µL in 384-well assay plates using the kit components at the recommended concentrations.

Sodium azide should **not** be added to the stock reagents. High concentrations of sodium azide (> 0.001 % final in the assay) might decrease the signal.

### Specific additional required reagents and materials:

The following materials are recommended:

Item	Suggested source	Catalog #
TopSeal-A PLUS Adhesive Sealing Film	PerkinElmer Inc.	6050185
VICTOR X, ViewLux, EnVision or EnSpire Multilabel Plate Reader equipped with TR-FRET option	PerkinElmer Inc.	-

### ○ Recommendations

#### General recommendations:

- The volume indicated on each tube is guaranteed for single pipetting. Multiple pipetting of the reagents may reduce the theoretical amount left in the tube.
- Centrifuge all tubes (including lyophilized analyte) before use to improve recovery of content (2000g, 10-15 sec).
- Re-suspend all reagents by vortexing before use.
- Use Milli-Q® grade H<sub>2</sub>O (18 MΩ•cm) to dilute Buffer.
- When diluting the standard or samples, change tips between each standard or sample dilution. When loading reagents in the assay microplate, change tips between each standard or sample addition and after each set of reagents.
- When reagents are added to the microplate, make sure the liquids are at the bottom of the well.
- Small volumes may be prone to evaporation. It is recommended to cover microplates with TopSeal-A Adhesive Sealing Films to reduce evaporation during incubation. LANCE *Ultra* TR-FRET assays cannot be read with the TopSeal-A Film attached. Please remove before reading.
- LANCE signal is detected using a VICTOR X, ViewLux, EnVision or EnSpire Multilabel Reader equipped with the TR-FRET. Use an excitation wavelength of 320 or 340 nm to excite the LANCE Europium chelate. We recommend you read this assay in dual emission mode, detecting both the emission from the Europium donor fluorophore at 615 nm, and the acceptor fluorophore (at 665 nm for *ULight* dye). The raw FRET signal at 665 nm can be used to process your data.
- Signal will vary with temperature and incubation time. For consistent results, identical incubation times and temperature should be used for each plate.
- The standard curves shown in this technical data sheet are provided for information only. A standard curve must be generated for each experiment. The standard curve should be performed in *Ultra* HiBlock Buffer

## ○ Assay Procedure

IMPORTANT: PLEASE READ THE RECOMMENDATIONS BELOW BEFORE USE

- The protocol described below is an **example** for generating one standard curve in a 20  $\mu\text{L}$  final assay volume (48 wells, triplicate determinations) and 452 samples. The protocols also include testing samples in 384 well plates. If different amounts of samples are tested, the volumes of all reagents must be adjusted accordingly, as shown in the table below. \*\*\*These calculations do not include excess reagents to account for losses during transfer of solutions or dead volumes.
- The standard dilution protocol is provided for information only. As needed, the number of replicates or the range of concentrations covered can be modified.
- Use of four background points in triplicate (12 wells) is recommended when LDL/LLOQ is calculated. One background point in triplicate (3 wells) can be used when LDL/LLOQ is not calculated.

Format	# of data points	Volume				Plate recommendation
		Final	Sample	Eu-Antibody	ULight Antibody	
TRF1280C	250	40 $\mu\text{L}$	30 $\mu\text{L}$	5 $\mu\text{L}$	5 $\mu\text{L}$	White OptiPlate-96 (cat # 6005290) White 1/2 AreaPlate-96 (cat # 6005560)
	500	20 $\mu\text{L}$	15 $\mu\text{L}$	2.5 $\mu\text{L}$	2.5 $\mu\text{L}$	White 1/2 AreaPlate-96 (cat # 6005560) White OptiPlate-384 (cat # 6007290)
	1 250	8 $\mu\text{L}$	6 $\mu\text{L}$	1 $\mu\text{L}$	1 $\mu\text{L}$	ProxiPlate™-384 Plus (cat # 6008280) White OptiPlate-384 (cat # 6007290)
	2 500	4 $\mu\text{L}$	3 $\mu\text{L}$	0.5 $\mu\text{L}$	0.5 $\mu\text{L}$	White OptiPlate-1536 (cat # 6004290)
TRF1280M	5 000	20 $\mu\text{L}$	15 $\mu\text{L}$	2.5 $\mu\text{L}$	2.5 $\mu\text{L}$	White 1/2 AreaPlate-96 (cat # 6005560) White OptiPlate-384 (cat # 6007290)
	12 500	8 $\mu\text{L}$	6 $\mu\text{L}$	1 $\mu\text{L}$	1 $\mu\text{L}$	ProxiPlate-384 Plus (cat # 6008280) White OptiPlate-384 (cat # 6007290)
	25 000	4 $\mu\text{L}$	3 $\mu\text{L}$	0.5 $\mu\text{L}$	0.5 $\mu\text{L}$	White OptiPlate-1536 (cat # 6004290)

**General Protocol (2-step protocol):** Dilute standards, samples, and assay components in 1X *Ultra* HiBlock Buffer.

**Each protocol described below is designed for 500 assay points including one standard curve (48 wells) and samples (452 wells).**

**Standard Preparation:**

1) Preparation of 1X *Ultra* HiBlock Buffer:

- a. Add 2 mL of 5X *Ultra* HiBlock Buffer to 8 mL H<sub>2</sub>O.

2) Preparation of CXCL9/MIG analyte standard dilutions:

- a. CXCL9/MIG analyte is provided at 0.3 µg in lyophilized form. Reconstitute with 100 µL H<sub>2</sub>O to create a 3 µg/mL solution. Prepare standard dilutions as follows (change tip between each standard dilution):

Tube	Vol. of CXCL9/MIG (µL)	Vol. of diluent (µL) *	[CXCL9/MIG] in standard curve	
			(g/mL in 15 µL)	(pg/mL in 15 µL)
A	10 µL of reconstituted CXCL9/MIG	90	3.00E-07	300 000
B	30 µL of tube A	60	1.00E-07	100 000
C	30 µL of tube B	70	3.00E-08	30 000
D	30 µL of tube C	60	1.00E-08	10 000
E	30 µL of tube D	70	3.00E-09	3 000
F	30 µL of tube E	60	1.00E-09	1 000
G	30 µL of tube F	70	3.00E-10	300
H	30 µL of tube G	60	1.00E-10	100
I	30 µL of tube H	70	3.00E-11	30
J	30 µL of tube I	60	1.00E-11	10
K	30 µL of tube J	70	3.00E-12	3
L	30 µL of tube K	60	1.00E-12	1
M ** (background)	0	100	0	0
N ** (background)	0	100	0	0
O ** (background)	0	100	0	0
P ** (background)	0	100	0	0

\* At low concentrations of analyte, a significant amount of analyte can bind to the vial. Therefore, load the analyte standard dilutions in the assay microplate within 60 minutes of preparation.

\*\* Four background points in triplicate (12 wells) are used when LDL is calculated. If LDL does not need to be calculated, one background point in triplicate can be used (3 wells).

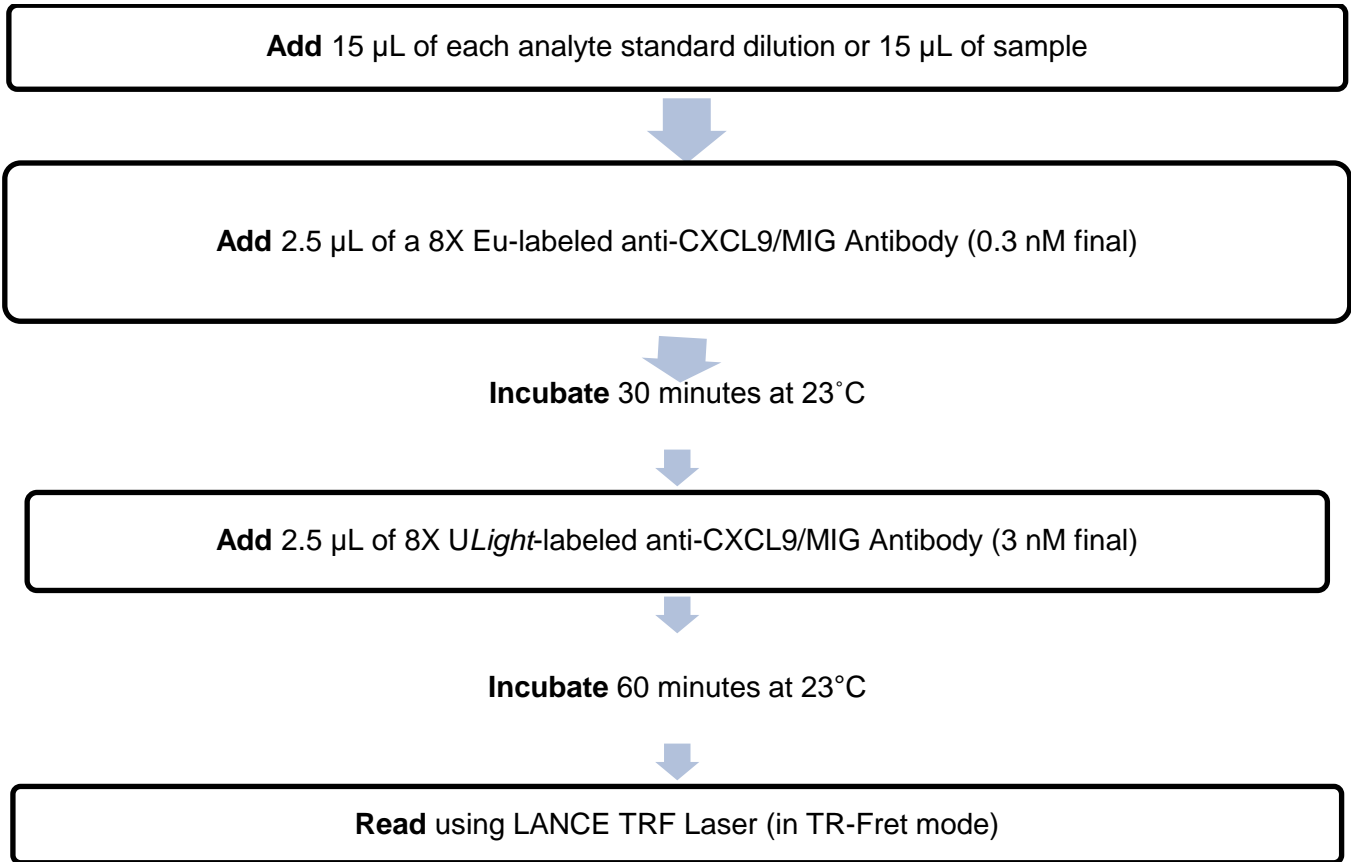
3) Preparation of 8X Eu-labeled anti-CXCL9/MIG Antibody (2.4 nM)

- a. Add 6 µL of 500 nM Eu-labeled anti-CXCL9/MIG Antibody to 1244 µL of 1X *Ultra* HiBlock Buffer.  
b. Prepare just before use.

4) Preparation of 8X ULight labeled anti-CXCL9/MIG Antibody (24 nM):

- a. Add 60 µL of 500 nM ULight labeled anti-CXCL9/MIG Antibody to 1190 µL of 1X *Ultra* HiBlock Buffer.  
b. Prepare just before use.

5) In a white Optiplate (384 wells):



## ○ Data Analysis

- Calculate the average count value for the background wells.
- Generate a standard curve by plotting the LANCE counts versus the concentration of analyte. A log scale can be used for either or both axes. No additional data transformation is required.
- Analyze data according to a nonlinear regression using the 4-parameter logistic equation (sigmoidal dose-response curve with variable slope) and a  $1/Y^2$  data weighting (the values at maximal concentrations of analyte after the hook point should be removed for correct analysis).
- The LDL is calculated by interpolating the average background counts (12 wells without analyte) + 3 x standard deviation value (average background counts + (3xSD)) on the standard curve.
- The LLOQ as measured here is calculated by interpolating the average background counts (12 wells without analyte) + 10 x standard deviation value (average background counts + (10xSD)) on the standard curve. Alternatively, the true LLOQ can be determined by spiking known concentrations of analyte in the matrix and measuring the percent recovery, and then determining the minimal amount of spiked analyte that can be quantified within a given limit (usually +/- 20% or 30% of the real concentration).
- Read from the standard curve the concentration of analyte contained in the samples.
- If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

## ○ Assay Performance Characteristics

*LANCE Ultra assay performance described below was determined using the 2 step protocol.*



### **Assay Sensitivity**

The LDL and LLOQ were calculated as described above. The values correspond to the lowest concentration of analyte that can be detected in a volume of 15 µL using the recommended assay conditions.

LDL (pg/mL)	LLOQ (pg/mL)	Buffer	# of experiments
11	56	<i>Ultra</i> HiBlock	9
7	42	DMEM	6
7	40	RPMI	6

### **Assay Precision:**

The following assay precision data were calculated from the three independent assays using two different kit lots. In each lot, the analytes were prepared in *Ultra* HiBlock Buffer. Each assay consisted of one standard curve comprising 12 data points in triplicate and 12 background wells containing no analyte. The assays were performed in a 384-well format using *Ultra* HiBlock Buffer.

#### **Intra-assay precision:**

The intra-assay precision was determined using 3 independent experiments for a total of 16 independent determinations in triplicate. CV% were calculated for each individual experiment then averaged. Shown is the average intra-experimental CV%.

CXCL9/MIG (CV%)	Buffer
2	<i>Ultra</i> HiBlock
2	DMEM
3	RPMI

#### **Inter-assay precision:**

The inter-assay precision was determined using the data across 3 independent experiments with 16 measurements in triplicate. CV% was calculated by comparing the same measurement in each experiment. The CV% for all 16 measurements were then averaged. Shown is the inter-experimental CV%.

CXCL9/MIG (CV%)	Buffer
11	<i>Ultra</i> HiBlock
10	DMEM
10	RPMI

**Spike Recovery:**

In four experiments, three known concentrations of CXCL9/MIG were spiked into 3 separate media and performed triplicate. The spiked samples were referenced to the CXCL9/MIG analyte curve produced in the corresponding media.

Spiked CXCL9/MIG (ng/mL)	% Recovery		
	Ultra HiBlock Buffer	DMEM + 10% FBS	RPMI + 10% FBS
3	89	102	96
1	91	97	91
0.3	95	118	108

○ **Troubleshooting Guide**

You will find detailed recommendations for common situations you might encounter with your LANCE *Ultra* Assay kit at:

<http://www.perkinelmer.com/Resources/TechnicalResources/ApplicationSupportKnowledgebase/LANCE/lance.xhtml>

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