

Dopamine D47 (RBHD47M400UA) saturation curve

Assay buffer:	50 mM Tris-HCl pH 7.4, 120 mM NaCl, 5 mM KCl, 5 mM MgCl ₂ , 1 mM EDTA		
Wash buffer:	50 mM Tris-HCl pH 7.4, 0.9 % NaCl		
Radioligand	[3H]-MethylSpiperone	85.5 Ci/mmol	1 µCi/1 µl
Cold ligand	Haloperidol		
Unit size	35 µg protein / unit		
Protein concentration	14 mg/ml		

Full scale assay format	
Total volume	550 µL
Volume membranes	500
Volume cold ligand or buffer	25
Volume radioligand	25

Membrane dilution 0.125ml membranes +
24.875 ml assay buffer
(1:200 dilution)

Haloperidol dilution Stock concentration 10 mM = 10 000 µM
Prepare 1000µL in assay
buffer
Final concentration 25 µM
Dilution factor 22
Intermediate concentration 550 µM

Volume haloperidol stock = $\frac{(1000 \mu\text{L} * 550 \mu\text{M})}{10\,000 \mu\text{M}}$ =55 µL stock

Radioligand dilution

Need 600 µL tube 1

Final Concentration 5 nM

Intermediate concentration 110 nM

$$\frac{110 \text{ nmol}}{1000 \text{ ml}} \times \frac{85.5 \text{ uCi}}{\text{nmol}} \times \frac{1 \text{ ul}}{1 \text{ uCi}} \times 0.6 \text{ ml} = 5.6 \text{ ul}$$

*Correction factor (1.3x) 5.6µL x 1.3 = **7.3µL**

Tube #	Final Concentration (nM)	Volume stock	Volume assay buffer
1	5	7.3 µl	593 µL
2	2	200 µl tube 1	300 µL
3	1	100 µL tube 1	400 µL
4	0.5	50 µL tube 1	450 µL
5	0.2	50 µL tube 2	450 µL
6	0.1	50 µL tube 3	450 µL
7	0.05	50 µL tube 4	450 µL
8	0.02	50 µL tube 5	450 µL

*Correction factor is used to account for ligand sticking to plastic; actual ligand concentration is later determined by spotting a set volume of working material from each tube onto a Filtermat, measuring dpm, and converting to moles using specific activity.

Plate map

Radioligand Concentration (nM)	Full Scale Saturation (550ul)					
	Total			Non-Specific		
	1	2	3	4	5	6
5	1	1	1	1	1	1
2	2	2	2	2	2	2
3	3	3	3	3	3	3
4	4	4	4	4	4	4
5	5	5	5	5	5	5
6	6	6	6	6	6	6
7	7	7	7	7	7	7
8	8	8	8	8	8	8

- 1) In DeepWell plate:
 - a. Add 25 µL buffer or Haloperidol
 - b. Add 25 µL radioligand
 - c. Add cell membranes (500 uL)
- 2) Incubate 1 hour
- 3) During incubation, pre-soak a UniFilter plate in 0.01% PEI
- 4) Wash assay plate over cell harvester, onto UniFilter plate
- 5) Dry UniFilter plate in 27°C oven
- 6) Add BackSeal to bottom of UniFilter plate
- 7) Add scintillation cocktail and TopSeal
- 8) Measure